

Inland Waters

ISSN: 2044-2041 (Print) 2044-205X (Online) Journal homepage: <http://www.tandfonline.com/loi/tinw20>

Spatial variation of nitrogen cycling in a subtropical stratified impoundment in southwest China, elucidated by nitrous oxide isotopomer and nitrate isotopes

Fu-Jun Yue, Si-Liang Li, Cong-Qiang Liu, Khan M.G. Mostofa, Naohiro Yoshida, Sakae Toyoda, Shi-Lu Wang, Shohei Hattori & Xiao-Long Liu

To cite this article: Fu-Jun Yue, Si-Liang Li, Cong-Qiang Liu, Khan M.G. Mostofa, Naohiro Yoshida, Sakae Toyoda, Shi-Lu Wang, Shohei Hattori & Xiao-Long Liu (2018) Spatial variation of nitrogen cycling in a subtropical stratified impoundment in southwest China, elucidated by nitrous oxide isotopomer and nitrate isotopes, Inland Waters, 8:2, 186-195, DOI: [10.1080/20442041.2018.1457847](http://www.tandfonline.com/action/showCitFormats?doi=10.1080/20442041.2018.1457847)

To link to this article: <https://doi.org/10.1080/20442041.2018.1457847>

Published online: 14 Jun 2018.

[Submit your article to this journal](http://www.tandfonline.com/action/authorSubmission?journalCode=tinw20&show=instructions) \mathbb{Z}

III Article views: 46

[View Crossmark data](http://crossmark.crossref.org/dialog/?doi=10.1080/20442041.2018.1457847&domain=pdf&date_stamp=2018-06-14)^で

Check for updates

Spatial variation of nitrogen cycling in a subtropical stratified impoundment in southwest China, elucidated by nitrous oxide isotopomer and nitrate isotopes

Fu-Jun Yue $\Phi,^{ab}$ Si-Liang Li,^{c,d} Cong-Qiang Liu,^a Khan M.G. Mostofa $\Phi,^d$ Naohiro Yoshida $\Phi,^e$ Sakae Toyoda,^e Shi-Lu Wang \bullet ,^a Shohei Hattori,^e and Xiao-Long Liu^f

^aState Key Laboratory of Environmental Geochemistry, Institute of Geochemistry, Chinese Academy of Sciences, Guiyang, China; ^bSchool of Geographical and Earth Sciences, University of Glasgow, Glasgow, UK; State Key laboratory of Hydraulic Engineering Simulation and Safety, Tianjin University, Tianjin, China; ^dInstitute of Surface-Earth System Science, Tianjin University, Tianjin, China; ^eDepartment of Environmental Science and Technology, Tokyo Institute of Technology, Yokohama, Japan; ^fTianjin Key Laboratory of Water Resources and Environment, Tianjin Normal University, Tianjin, China

ABSTRACT

Estimates of biogeochemical processes and the proportion of $N₂O$ production in the aquatic system of impoundments are important to quantify nitrogen cycling, particularly during stratification periods. In this study, we used the dual isotopes of nitrate $(NO₃)$ and nitrous oxide $(N₂O)$ to estimate the nitrogen dynamics and contributions to $N₂O$ production and reduction at varying zones in Lake Baihua, located in southwest China. The lake was strongly stratified during the sampling period, with the oxic zone from the surface to 12 m and the anoxic zone from 12 to 21 m. The assimilation shifted $\delta^{15}N$ and $\delta^{18}O$ of NO₃ significantly in the epilimnion (0–4 m), and denitrification contributed to the shift in the low dissolved oxygen zone of the hypolimnion. The semiquantitative analysis showed that nitrification accounted for $>67\%$ of the N₂O production between 0 and 4 m while higher nitrification contributions were also found between 6 and 12 m. The contribution of denitrification between 15 and 21 m was >43%. The mechanism responsible for the vertical variations should be considered in the estimation of nitrogen cycling and N_2O production in subtropical stratified impoundments.

Introduction

Lakes are considered a large global sink for reactive nitrogen (Pearson et al. [2012](#page-9-0), Finlay et al. [2013](#page-8-0)). Nitrogen (N) dynamics in lake ecosystems involve a series of complex processes, such as external inputs and internal biogeochemical cycles; however, the effects of freshwater N removal are still poorly understood (Mayer and Wassenaar [2012](#page-9-0), Finlay et al. [2013,](#page-8-0) Wenk et al. [2014\)](#page-9-0). In general, nitrate $(NO₃⁻)$ is the dominant form of dissolved inorganic nitrogen (DIN) entering aquatic environments. Globally, concentrations of $\overline{\text{NO}_3}^-$ are steadily increasing in aquatic systems, which has been attributed to increasing surface run-off, river damming, aquaculture, N deposition, agricultural, and other landbased activities (Sterner [2011](#page-9-0), Mostofa et al. [2013](#page-9-0), North et al. [2013](#page-9-0), Beusen et al. [2016](#page-8-0)). High levels of $NO₃⁻$ in freshwaters are detrimental to human and ecosystem health and sustainability (Ohte et al. [2010,](#page-9-0) Finlay et al. [2013](#page-8-0), Beusen et al. [2016](#page-8-0)).

Lakes are most likely to become important N sinks under stratified conditions, especially for anoxic bottom water. Conditions in the surficial bottom sediments of lakes that favor denitrification processes are due to high organic matter inputs from the overlying water column and strong dissolved oxygen (DO) gradients. Denitrification processes in stratified lakes primarily occur in the redox transition zone, which is largely controlled by the oxygen supply via hydrodynamic processes and the consumption of oxidants due to organic matter degradation (Pearson et al. [2012,](#page-9-0) Wenk et al. [2014\)](#page-9-0). Nitrous oxide (N_2O) can be produced by the N cycle as a by-product of hydroxylamine $(NH₂OH)$ oxidation during nitrification processes as well as during incomplete denitrification processes (Sasaki et al. [2011,](#page-9-0) Wenk et al. [2016\)](#page-9-0) and can contribute to global warming and stratospheric ozone depletion (Well et al. [2005\)](#page-9-0).

Stable isotope analysis, particularly the dual isotope approach of NO_3^- and N_2O , is a powerful tool that can provide critical insights into N transformation dynamics in lakes (Spoelstra et al. [2001,](#page-9-0) Xiao and Liu [2004](#page-9-0), Finlay et al. [2007,](#page-8-0) Ohte et al. [2010](#page-9-0), Sasaki et al. [2011,](#page-9-0) Mayer and Wassenaar [2012](#page-9-0), Wenk et al. [2014,](#page-9-0) [2016\)](#page-9-0). For example, the values of dual $NO₃$ isotopes would increase during biological processes in aquatic environments, such as

ARTICLE HISTORY Received 18 July 2017 Accepted 1 February 2018

KEYWORDS denitrification; N₂O isotopomer; nitrate isotopes; nitrification; stratified lake

uptake by algae or denitrification by bacteria. Previous studies showed that the $\delta^{15}N$ and $\delta^{18}O$ isotopic ratio was maintained at 1:1 during assimilation (Granger et al. [2010,](#page-9-0) Mayer and Wassenaar [2012](#page-9-0)) but increased to ∼2:1 during denitrification (Mayer and Wassenaar [2012](#page-9-0), Wenk et al. [2014](#page-9-0)).

The ¹⁵N-site preference (SP), namely the difference in isotopic ¹⁵N between the central (α) and terminal (β) positions within the asymmetric N_2O molecule, can be defined as $SP = \delta^{15}N^{\alpha} - \delta^{15}N^{\beta}$ (Toyoda and Yoshida [1999](#page-9-0)). The characteristic isotopic signature of N_2O can be used to determine different sources of N_2O and its microbial pathways (Yoshida et al. [1989](#page-10-0), Toyoda et al. [2005](#page-9-0), Sutka et al. [2006](#page-9-0), Yamagishi et al. [2007](#page-9-0), Wenk et al. [2016\)](#page-9-0). Previous studies have used SP as a diagnostic tool to elucidate source processes of N_2O , such as nitrification and denitrification (Sutka et al. [2003](#page-9-0), Baggs [2008](#page-8-0), Frame and Casciotti [2010](#page-8-0), Santoro et al. [2011](#page-9-0), Heil et al. [2014](#page-9-0)). For example, the SP signature of N_2O via nitrite $(NO₂⁻)$ or $NO₃⁻$ reduction, whether by nitrifying or denitrifying bacteria, can be as low as −10.7‰, clearly lower than that from N_2O produced by nitrification (e.g., ammonia and hydroxylamine oxidation, up to 37.5‰; Sutka et al. [2006,](#page-9-0) Frame and Casciotti [2010\)](#page-8-0). In addition, fungal denitrification yields N_2O with an SP value of 37‰, which is close to nitrification but considerably different from that produced during bacterial denitrification (Sutka et al. [2008\)](#page-9-0). Finally, pure culture studies provide compelling evidence suggesting that SP values of 0‰ or less are indicative of N_2O produced by denitrifying bacteria while SP values of 30.6‰–7.5‰ indicate N2O produced from hydroxylamine oxidation or fungal denitrification (Sutka et al. [2006,](#page-9-0) Frame and Casciotti [2010](#page-8-0), Wu et al. [2016\)](#page-9-0).

Hydroelectric impoundments are widely distributed in southwest (SW) China. High levels of N_2O and its emission to the atmosphere have been found from those reservoirs (Liu et al. [2011](#page-9-0)). Constructed in 1966, Lake Baihua (BH) is a typical hydroelectric impoundment in SW China with a series of 8 dams along the Maotiao River, located in the upper reaches of Wujiang, a tributary of the Changjiang (Yangtze) River of China. BH is a eutrophic lake with a high content of dissolved organic matter (up to 330 μM C) and high primary production with chlorophyll concentration up to 66 μg/L (Fu et al. [2010\)](#page-9-0). Previous studies showed that the production and consumption of $N₂O$ in BH strongly accord with the decomposition of organic matter during lake stratification, and the emission flux of N_2O is high (2.4 μmolm2 /h; Wang et al. [2004](#page-9-0), Liu [2010\)](#page-9-0). However, the proportion of N_2O production from various processes is not well constrained in hydroelectric impoundments of SW China. Relatively high concentrations of $NO₃$

and $N₂O$ along with high contents of dissolved organic matter in BH make it ideal for studying N processes in subtropical impoundments, which can provide vital information for understanding processes for many lakes with similar conditions around the globe. To understand the processes leading to the production of N_2O , however, it is imperative to explore the N cycle in the whole water column. As such, the goal of this study was to assess NO_3^- and N_2O transformation in this subtropical lake during summer stratification periods. We report new measurements to estimate the concentration and isotopic composition of $NO₃$ and $N₂O$ in the water column to assess their spatial variation as well as to identify their fate. The major transformations of nitrification and denitrification contributed to N2O production and reduction in different water depths were also estimated based on SP values.

Study site

BH is located near Guiyang, SW China, at an altitude of ∼1200 m a.s.l. The mean annual precipitation in the area is ∼1200 mm. The valleys upstream and downstream of BH are both long and narrow, but the middle section of the lake is wide, with >100 islands distributed in the lake (Fig. 1). Another hydroelectric impoundment, Lake Hongfeng (HF) built in 1960, is situated upstream of BH. The water discharged from HF (30.2 m^3/s) is the primary water source for BH $(38 \text{ m}^3/\text{s}; \text{ Liu } 2010)$ $(38 \text{ m}^3/\text{s}; \text{ Liu } 2010)$. These 2 impoundments serve to control floods and provide water for electrical power generation, industrial and

Figure 1. Lake Baihua sampling sites, denoted by circles. The gray areas within the lake represent islands.

agricultural water supply, and recreation. The water level fluctuates between 1240 m (normal level) and 1227.5 m (storage level) a.s.l. in HF, and between 1195 m and 1188 m a.s.l. in BH, influenced by engineered water control and climate (Liu [2010\)](#page-9-0). Stratification can be observed frequently between April and August in BH (Wang et al. [2004](#page-9-0)).

Methods

Water sampling was conducted in BH during August 2013. To understand the spatial variations in N cycles within the entire lake system, 4 sampling sites were chosen along the lake [\(Fig. 1](#page-2-0)). Specifically, the first sampling site $(BH₁)$ was located at the upper reach, the second $(BH₂)$ was near one of the tributaries located in the middle section, the third $(BH₃)$ was located in the lower reach, and the fourth (BH4) was located ∼0.5 km upstream of the dam. Water flow was from $BH₁$ to BH₄ ([Fig. 1](#page-2-0)). Profiles of temperature (T), DO, and conductivity (EC) were obtained with an automated multiparameter monitoring instrument (Yellow Springs Instruments: YSI 6600 v2). A 5-L Niskin water sampler was used to collect water samples at the same depths at which T, DO, and EC were measured. Water samples were filtered through 0.45 μm membrane filters (Millipore) and kept at 4 °C in the dark until analysis. Concentrations of ammonium (NH₄), NO₂, NO₃, and total dissolved nitrogen for the filtered samples were analyzed using an automatic flow analyzer (SKALAR Sans Plus Systems) within 48 h. The concentrations of dissolved organic carbon (DOC) were analyzed as $CO₂$ by the wet oxidation method (Aurora 1030). Water samples were first acidified with 5% (vol/vol) of H_3PO_4 to remove all inorganic carbon, and then 10% $Na₂S₂O₈$ was added to oxidize organic carbon to inorganic carbon.

The $NO₃⁻$ isotopes of water samples were measured by the denitrifer method (McIlvin and Casciotti [2011\)](#page-9-0). Four international NO_3^- (USGS-32, USGS-34, USGS-35, and IAEA-N3) and 2 experimental reference materials were used for the calibration. The precision for the samples analyzed in duplicates was 0.3‰ for $\delta^{15}N$ and 0.5‰ for $\delta^{18}O$ of NO₃.

Water samples for N_2O isotopomer analysis were directly sampled into 120 mL glass bottles sealed with butyl rubber stoppers. The samples were injected with 0.6 mL of HgCl₂ solution (5%) and stored at 4 $^{\circ}$ C until analysis. Bulk N_2O isotope and isotopomer analyses were conducted at the Tokyo Institute of Technology, Japan, using an online analytical system with a stainless steel gas transfer line, pre-concentration traps, chemical traps for removal of H_2O and CO_2 , and a gas chromatograph/isotope ratio mass spectrometer (MAT 252; Thermo Fisher Scientific Inc.; Toyoda and Yoshida [2016\)](#page-9-0). The detailed description of the calibrated method and process for measuring $N₂O$ concentrations and isotopomer can be found in Toyoda and Yoshida [\(2016\)](#page-9-0). The $\delta^{15}N$ and $\delta^{18}O$ were reported relative to atmospheric N₂ and V-SMOW, respectively.

Results

Because of the shallow depth at $BH₁$, the depth profiles of T, DO, and DOC (Fig. 2) were limited to 10 m. The T profiles in the epilimnion zone were similar at all 4 sites (Fig. 2a); however, below the thermocline (∼6–10 m), the depth-profiles of T remained similar between the 2 middle sites (i.e., $BH₂$ and $BH₃$) but were notably different at the other 2 sites. The mixing discharge at site $BH₄$, with a constant T value between 15 and 21 m, may be caused by continuous water flow at the suboxic hypolimnion zone toward the dam. The DO values ranged from 223 to 338 μmol/L (μM) from the surface to the depth of 4 m and then decreased rapidly between the depth of 4 and 8 m (Fig. 2b). Low DO (<20 μ M) was detected at 10 m at BH₁ and below 12 m at the other 3 sites. Thus, the oxic zone extended from the surface to about 12 m, and the anoxic zone was roughly from 12 m to the lake bottom. In addition, DO values between the surface and 6 m were higher at the upper section of the lake (i.e., $BH₁$) and gradually decreased toward the lower reaches (i.e., $BH₄$). This decreasing trend in DO along the water flow direction was probably caused by O_2 degassing and consumption of oxygen by degradation of organic matter. As the major recharge source of BH, the water discharged from HF with low T (13.7 °C) can increase dissolution of O_2 into the water, but the increased temperature of water during the water flow may lead to O_2 degassing. Although DOC concentrations were relatively constant

Figure 2. Water column profiles of (a) temperature, (b) DO, and (c) dissoved organic carbon in Lake Baihua during August 2013.

Figure 3. Water column profiles of (a) dissolved NO₃, (b) DON, (c) NH¹4, (d) NO₂, (e) δ^{15} N–NO₃, and (f) δ^{18} O–NO₃ in Lake Baihua during
August 2013 August 2013.

at BH₄, they varied greatly with depth at BH₂ and BH₃, where higher DOC concentrations were found at deeper depths ([Fig. 2c](#page-3-0)).

The depth profiles of NO_3^- , dissolved organic nitrogen (DON), NH_4^+ , NO_2^- , and dual NO_3^- isotopes at the 4 sampling sites (Fig. 3) show that the average concentration of $NO₃⁻$ over the depth profile was highest at BH₁ (mean [1 SD] = 119 [24] μM) and decreased in the lower reaches with (86 [47] μ M) at BH₂, (81 [35] μM) at BH₃, and (85 [17] μM) at BH₄ (Fig. 3a). NO₃ concentrations were relatively constant above 4 m, but in the deeper water its concentrations fluctuated greatly with depth and among sites (Fig. 3a).

The average concentration of DON over the depth profile was lower at $BH₁$ (10.8 [5.5] μ M) and increased monotonically from $BH₂$ (15.9 [8.0] μ M) to $BH₃$ (23.4 [27.6] μM) and BH₄ (33.3 [12.3] μM; Fig. 3b). DON at BH₂ showed the greatest fluctuation with depth among all sampling sites, with higher values at 4, 12, and 21 m. Significant negative relationships between DON and $NO₃⁻$ existed at all sites except for $BH₂$, which were indicative of the contribution of $NO₃$ from degradation of organic N (Fig. 4a).

 NH_4^+ and NO_2^- were minor forms of DIN while $NO_3^$ and DON were the dominant forms of dissolved N (Fig. 3c). The concentrations of NH_4^+ and NO_2^- were low (<4 μ M) for most depth profiles. The NH⁺ concentration was highest at 6 m at $BH₁$ and 2 m at $BH₄$ (Fig. 3c), and $NO₂⁻$ concentration was highest at 12 m at $BH₂$ (Fig. 3d).

The isotopic composition of $NO₃⁻$ varied markedly with depth at all 4 sites (Fig. 3e). The average $\delta^{15}N$ of NO₃ over the depth profile increased from 15.2‰ (2.4‰) at BH_1 to 20.3‰ (5.1‰) at BH_2 and 23.0‰ (9.1‰) at $BH₃$ and then decreased slightly to 21.8‰ (5.8‰) at BH₄. The average $\delta^{18}O$ of NO₃ over the

Figure 4. The relationship between (a) NO₃ and DON and between (b) δ^{15} N-NO₃ and δ^{18} O-NO₃ in Lake Baihua during the summer stratified period stratified period.

depth profile varied from 6.7% (1.6‰) at BH₁ to 6.8% (6.1%) at BH₂, 7.8‰ (5.1‰) at BH₃, and 6.1‰ (2.9‰) at $\rm BH_4$. The $\rm NO_3^-$ isotope peaks occurred at the top of the anoxic zone (between 4 and 6 m). However, $\delta^{18}O$ of $NO₃⁻$ tended to decrease with depth, and negative values were measured at 15 and 18 m at $BH₂$ and $BH₃$, respectively. Increasing $\delta^{15}N$ and $\delta^{18}O$ of NO_3^- with decreasing NO₃ concentrations were also observed in a few water samples taken from the epilimnion and in most of the water samples taken from the hypoxic hypolimnion. The highest $\delta^{15}N$ value was 40.5‰ at 6 m at BH₃ and the highest $\delta^{18}O$ value was also observed at the same depth at $BH₃$ with a value of 16.9‰. In addition, the $\delta^{15}N$ and $\delta^{18}O$ values of NO₃ were relatively stable from 0 to 8 m at $BH₁$ and below 12 m at $BH₄$, whereas those values fluctuated at $BH₂$ and $BH₃$, closely resembling the trends in NO_3^- and DON concentration profiles. A 1:1 slope line existed between $\delta^{18}O$ of NO₃ and $\delta^{15}N$ of $NO₃⁻$ for the water samples taken from the depths between 0 and 4 m ([Fig. 4b\)](#page-4-0).

The $N₂O$ concentrations varied substantially among the sampling sites ranging from 15 to 850 nmol/L (nM), and the average values increased from 33.9 (27.4) nM at BH₁, to 77.1 (81.4) nM at BH₂, to 213 (274) nM at $BH₃$ and then decreased to 45.4 (27.5) nM at BH4 (Fig. 5a). Extreme oversaturation levels (e.g., 1.6–71-fold) of $N₂O$ with respect to atmospheric equilibrium were detected at different depths. Three water profiles had N_2O peaks at 8 m, associated with the rapid decrease in DO concentrations (Fig. 5a and [2b\)](#page-3-0). Although the N_2O peak at BH_4 occurred at 11 m, it occurred at analogous DO concentrations to other depth profiles with high concentrations of N_2O (Fig. 5a and [2b\)](#page-3-0) and was associated with the lowest bulk $\delta^{15}N$ (−8.3‰). The highest N₂O concentration was detected at the bottom of $BH₃$, which also had the highest DOC concentration. The stable isotopic composition of N_2O at $BH₁$ was relatively constant with depth, except that a peak value occurred at 8 m, which corresponded to the depth of the peak concentration of N_2O (Fig. 5a–b).

The bulk $\delta^{15}N$ of N₂O ranged from -8.3% to 26.9‰ for the 4 water depth profiles. The average values of bulk δ^{15} N were 2.8‰ (1.8‰) at BH₁, 6.8‰ (8.7‰) at BH₂, 2.0‰ (2.9‰) at BH₃, and 2.8‰ (9.2‰) at BH₄. The bulk δ^{18} O values of N₂O ranged from 16.1‰ to 55.1‰ among the 4 depth profiles, and their average values were 26.7‰ (5.3‰) at BH₁, 36.8‰ (6.8‰) at BH₂, 31.8‰ (6.0‰) at BH₃, and 32.9‰ (14.2‰) at BH₄. The bulk $\delta^{15}N$ and $\delta^{18}O$ were higher at 12 m and then decreased to the bottom at $BH₂$; however, the bulk δ^{15} N decreased from 8 m to the bottom and δ^{18} O decreased from 12 m to the bottom at BH₃.

The average SP values of N_2O were higher in the upper $(29.2\% \text{ } [11.1\% \text{] at } BH_1)$ and lower $(33.0\% \text{)}$ $[16.8\%]$ at BH₄) reaches compared to the 2 middle sites (26.9‰ [9.0‰] at BH₂ and 20.4‰ [9.7‰] at BH₃). The SP values were higher at 8 and 15 m and subsequently decreased to the bottom for 3 profiles.

Discussion

Nitrate transformation and cycling along with its isotopic characteristics in Lake Baihua

Compared to other sites, $BH₁$ showed higher $NO₃⁻$ concentrations and lower NO_3^- isotopes at almost all depths, indicating different NO_3^- sources or different processes. Degradation of organic N (e.g., organic N \rightarrow NH₄⁺ \rightarrow NO₃; mineralization and nitrification), which is primarily released from phytoplankton by both photochemical

Figure 5. Water column profiles of (a) N₂O, (b) $\delta^{15}N_{bulk}-N_2O$, (c) $\delta^{18}O-N_2O$, and (d) SP-N₂O in Lake Baihua during August 2013.

and microbial respiration, can produce $NH₄⁺$ and then NO3 – . Significant negative relationships between DON and NO_3^- at all sites except BH_2 were indicative of the contribution of NO_3^- from degradation of organic N ([Fig. 4a](#page-4-0)). The isotopic fractionation in these paths is likely to cause the nitrified NO_3^- to have a lower isotopic signature than that of the primary sources. The terrestrial inputs and high DON concentration in drainage water from the bottom of the lake might result in higher $NO₃⁻$ concentrations at BH₁ than at the other 3 sites.

At the 3 other sites, $NO₃⁻$ isotopes decreased from 12 m to the bottom of the lake subsequent with increased DON concentration. The high concentrations of DON at greater depths might be the result of phytoplankton sink-ing out of surface waters (Mostofa et al. [2016\)](#page-9-0). Low $\delta^{18}O$ - $NO₃$ values might be caused by the oxygen exchange between water and $NO₂⁻$ during nitrification or by the different isotopic value of water oxygen and atmospheric oxygen used during nitrification (Casciotti et al. [2010\)](#page-8-0). N characteristics in deeper water layers (>12 m) suggested that NO_3^- at those depths was primarily derived from nitrification. Furthermore, dissimilation and denitrification processes may be also involved in the 3 depth profiles and result in higher NO₃ isotopes. The 1:1 line between δ^{18} O of NO₃ and δ^{15} N of NO₃ in the 0–4 m depth samples would represent assimilation along with the regeneration of NO_3^- via nitrification ([Fig. 4a](#page-4-0); Granger et al. [2010](#page-9-0), Mayer and Wassenaar [2012](#page-9-0)).

Although mineralization and nitrification were the major N-transformations at $BH₁$, the NO₃ characteristics at the bottom of $BH₁$ also indicated incomplete denitrification. The decrease in $NO₃⁻$ concentration from 12 m to the bottom of the lake suggested dissimilation or denitrification processes. As one of the 2 reductive N reduction pathways, however, dissimilation mostly occurred in the epilimnion. More specifically, this decrease in $NO₃⁻$ concentrations with depth was caused by denitrification. The mixing between denitrification and nitrification was the major reason for the vertical variation in the NO_3^- concentration from 12 m to the bottom of the lake, subsequently with relatively low NO₃ concentration and isotopes and high DON.

$N₂O$ cycling along with its isotopic characteristics in Lake Baihua

The $N₂O$ concentration and its isotopic composition were the net result of mixing dissolved N_2O from the upper reaches (drainage water from the bottom of HF) with simultaneous production and reduction of N_2O . N_2O reduction has been thought to be more affected by low oxygen levels than other enzyme-regulated steps involved in denitrification. Moreover, enzyme-regulated steps could result in a lag phase between reduction and production of N_2O , which could possibly lead to a shortterm accumulation of N_2O (Wenk et al. [2016\)](#page-9-0). High fluctuations of N_2O concentration in the oxic zone, oxic–anoxic interface, and the anoxic water at depth may be linked to enhanced denitrification rates and subsequent regulation of reduction (stimulated by increased organic matter inputs following spring algal blooms).

The SP values of N_2O varying between -1.5% and 47.7‰ showed a slight variation in the epilimnion and a more pronounced variation in the hypolimnion, the latter of which was larger than those for other lakes (Sasaki et al. [2011](#page-9-0), Wenk et al. [2016\)](#page-9-0). The simultaneous increases in NO_3^- and N_2O concentrations between the surface and 8 m in these 3 profiles, except for at 6 m at $BH₃$, indicated $N₂O$ production by nitrification. Some features can be obviously identified for NO_3^- and N2O. For example, high SP values were found at 4 and 8 m at $BH₂$ and at 8 m at $BH₃$ with the N₂O peak and high SP value, which was interpreted as the result of nitrification processes and notably different from at 6 m at BH₃. The N₂O concentration at BH₄ was highest at 11 m, where a minimum bulk $\delta^{15}N-N_2O$ (i.e., -8.3%) occurred, indicating that *in situ* N₂O production could occur by either bacterial nitrification or nitrifier denitrification. The SP value was 43.6‰ at the depth of N_2O production, however, similar to the N_2O production at 8 m at $BH₃$, suggesting an $NH₂OH$ -derived source of N_2O rather than nitrifier denitrification. Nitrification was observed for all profiles with the highest peaks for both NO_3^- and N_2O concentrations at 8 m, suggesting a simultaneous occurrence of $NO₃$ regeneration and N_2O production by nitrifying microorganisms. The amount of newly produced N_2O was expected to be controlled by the newly produced $NO₃$, if nitrification was related to the N_2O accumulation in the oxic zone.

Although the isotopic evidence implied that nitrification was responsible for the increased SP values, samples at 6 m at \overline{BH}_3 with considerably increased NO₃ isotope values and decreased SP values could indicate that $\overline{NO_3}$ and N_2O mainly originated from denitrification processes. Denitrification was also observed in the anoxic hypolimnion (15 to 21 m) zone with decreased SP signatures and $NO₃⁻$ concentrations, suggesting the increased contribution of denitrification to N_2O production. The highest DOC concentration observed at the bottom of $BH₃$ and anoxic zones was conducive to denitrification. Low values of the N₂O isotopomer (SP = -1.5% and $\delta^{15}N_{bulk} = -2.1\%$ at 21 m at BH₃ indicated that denitrification was responsible for the highest N_2O concentrations at this depth.

Elucidating the oxygen isotopic characteristics of N_2O is particularly challenging because its isotopic composition reflects both source and the tendency for intermediate compounds of N_2O production (Sutka et al. [2006,](#page-9-0) Well and Flessa [2008](#page-9-0), Zou et al. [2014](#page-10-0)). The δ^{18} O of N₂O derived from NH₂OH decomposition might depend on the δ^{18} O of O₂ during the first reaction of nitrification if the δ^{18} O of N₂O produced by nitrification and nitrifier–denitrification depended on both the $\delta^{18}O-O_2$ and $\delta^{18}O-H_2O$, which would be proportionally affected by the amount of oxygen atom exchange between $NO₂$ and $H₂O$. This pattern is consistent with relatively more N_2O production by nitrifier-denitrification due to a significant decrease in $O₂$ concentration at deeper layers; thereby, a substantial contribution to the overall $\delta^{18}O-N_2O$ came from H₂O.

The contribution of nitrification, denitrification, and reduction to N_2O

Relatively stable N_2O concentrations and SP values between the surface and 4 m at $BH₁$, $BH₂$, and $BH₃$ implied the homogeneous distribution of N_2O . The dissolved N_2O in the upper reaches might partly come from denitrification owing to the outflow from the bottom of HF. The SP signatures at those depths suggested that N_2O between the surface and 4 m was a mixture of denitrified N_2O from the upper reaches and nitrified N_2O in situ. The discrepancy of SP signatures between the 2 production pathways can be used to back-calculate the relative contributions of nitrification and denitrification to the $N₂O$ production according to the following equation:

$$
\delta_{measure} = \delta_{denitrification} \times (1 - f) + \delta_{nitrification} \times f, \qquad (1)
$$

where $\delta_{\text{denitrification}}$, $\delta_{\text{nitrification}}$, and f are denoted as the SP value for denitrification, the SP value for nitrification, and the nitrification contribution, respectively.

To further quantify the contributions of the 2 processes, the SP signature of endmembers should be determined. In this study, the high concentration of N_2O in the deep water (21 m) at the center of the lake (BH_3) suggested that denitrification was responsible for the accumulation. The low values of N_2O isotopomers and the highest N_2O concentrations in the deep water (21 m, $SP = -1.5\%$) at the center of the lake (BH₃), which was within the range of pure cultures from −4.0‰ to 1.9‰ (Stuka et al. 2006), can be used as a denitrification signature because of the relatively stable stratification. The SP value of hydroxylamine oxidation or nitrification from pure cultures ranged from 30.6‰ to 37.5‰, and the SP value of 33‰ was characteristic of nitrification (Stuka et al. 2006). We used an SP of −1.5‰ and 33‰ as the characteristic value of denitrification and nitrification, respectively. The contribution of $N₂O$ production by nitrification and denitrification at BH₄ was not calculated because this site location was close to the dam and the trend of N_2O isotopic composition significantly varied compared to the other 3 sites. The contribution of nitrification in the epilimnion zone between the surface and 4 m (SP 20.2−27.8‰) was calculated to be between 70% (6.8%), 75% (9.3%), and 67% (5.8%) at BH₁, BH₂, and BH₃, respectively.

As discussed earlier, the relatively low SP value in the hypolimnion zone (15 to 21 m) can be explained by a mixture of nitrification and denitrification processes. Using the same end member information, the portion of the denitrification contribution can be calculated. The results showed that 42% (3.6%) and 57% (41%) of N_2O were produced by denitrification at BH_2 and BH_3 , respectively, whereas 58% (3.6%) and 43% (41%) of N_2O were produced by nitrification at BH₂ and BH₃, respectively. Because the SP value at 21 m at $BH₃$ was chosen as the end member of denitrification, the denitrification contribution should be responsible for the entire N₂O production at this depth.

From 6 to 12 m, oxygen concentrations decreased with depth. This decrease may shift the N_2O production process from nitrification to denitrification and also the increase in the ratio of $N₂O$ reduction to its production, which may induce an increase in SP value along with $\delta^{15}N_{N2O}$ and $\delta^{18}O_{N2O}$ values (Well et al. [2005](#page-9-0), Yamagishi et al. [2007](#page-9-0), Koba et al. [2009,](#page-9-0) Wenk et al. [2016\)](#page-9-0). From the various reports of high SP values in aquatic systems, high SP values were characteristic of denitrifying aquatic environments, where N_2O reduction is favored because N_2O fluxes are restricted by low gas diffusivity (Well et al. [2005\)](#page-9-0). In this study, no profiles showed a consistent increase in δ^{15} N, δ^{18} O, and SP values. Although higher SP values than the pure culture results were observed, N_2O reduction can be used to explore possible mechanisms for the high values. N_2O between 6 and 12 m likely resulted from mixing processes, including nitrification and denitrification, and the increase in the ratio of N_2O reduction. If the contribution of nitrification and denitrification between 6 and 12 m was the same as that between the surface and 4 m, the SP signature should have the same range as the signature between the surface and 4 m (SP 20.2–27.8‰). The proportion of N_2O reduction can be calculated using the following approximate Rayleigh linearization (Yamagishi et al. [2007](#page-9-0), Sasaki et al. [2011\)](#page-9-0):

$$
\delta_{\text{measure}} = \delta_{\text{original}} + \epsilon \times \ln(C/C_0), \qquad \qquad (2)
$$

where $\varepsilon = {}^{SP} \varepsilon = -16.4\%$, which is calculated from the N2O reduction in the water column (Yamagishi et al. [2007,](#page-9-0) Sasaki et al. [2011](#page-9-0)), and $\delta_{original} = SP_{original} =$ 22.9‰, which is the average SP signature between the

surface and 4 m at $BH₁$, $BH₂$, and $BH₃$. $C₀$ and C denoted the original, not reduced, N_2O concentration and N_2O concentration after reduction, respectively. Quantitative analysis showed that the pre-existing N_2O could be reduced within the range of 24% to 77% at $BH₂$ and $BH₃$.

Although we calculated the N_2O contributions from nitrification, denitrification, and reduction based on the mixing model and Rayleigh linearization, uncertainties might still exist in those calculations. For example, unstable redox conditions between 6 and 12 m might impede the transformation of N_2O to N_2 (Wenk et al. [2016](#page-9-0)). Thus, the N_2O reduction might be overestimated between 6 and 12 m because the SP signature of nitrification can reach up to 37.5% (Frame and Casciotti 2010). The high SP signature implied that nitrification contributions between 6 and 12 m should be higher than between the surface and 4 m. Furthermore, we could not exclude the possibility of contribution of N_2O reduction between 15 and 21 m, meaning that the quantitative analysis from denitrification between 15 and 21 m might be underestimated. In addition, this calculation ignored the vertical and horizontal diffusion, which might influence the estimation based on the isotopic fractionation (Well and Flessa [2008](#page-9-0)).

Conclusion

This study investigated N biogeochemical processes and N2O production in a typical stratified impoundment in SW China, located on a river with a large number of hydroelectric dams. The water temperature profile showed the thermocline occurred between a depth of 6 and 8 m. The DO concentration declined in the thermocline and continued to fall to <20 μM below 12 m. The DOC concentrations showed a larger variation, particularly at $BH₂$, whereas high concentrations of DOC were detected at the bottom of the other 3 profiles. $NO₃⁻$ concentrations and its isotopic composition varied with depth and among locations. The positive relationship between $\delta^{15}N$ and $\delta^{18}O$ of NO₃ along with decreasing NO₃ in the 0–4 m depth could be a signature of NO₃ assimilation. Denitrification and nitrification were responsible for the variation of various N species from 12 m to the bottom of the lake.

High N_2O concentration and its extreme oversaturation were often detected for all depth profiles of Lake BH. Simultaneous increases in NO_3^- and N_2O concentrations at $BH₁$ indicated that the N₂O production could result from nitrification. The detected N_2O peaks suggested N_2O accumulation. The SP signature suggested that N_2O in the oxic zone was produced by denitrified N_2O from the upper reach and nitrified $N₂O$ *in situ*. Low values of the $N₂O$ isotopomers along with the highest N_2O concentrations at the bottom of this lake $(SP = -1.5\%)$ can be used as a signature for denitrification. Although the calculation of SP gave semiquantitative analysis for N_2O dynamic processes, we concluded that nitrification contributed >67% of the N2O production between the surface and 4 m, which was lower than that between 6 m and 12 m. N_2O reduction is <77% between 6 and 12 m. The contribution of denitrification between 15 and 21 m was >43%. Further studies on the vertical distribution of microorganism and hydrological conditions in the key zone will help to understand N dynamics and production of greenhouse gases in hydroelectric impoundments.

Acknowledgements

This work was supported by the National Key R&D Program of China under Grant No. 2016YFA0601002; National Natural Science Foundation of China under Grant No.s 41571130072, 41403105, 41672351; and IAEA under Grant F32007. We thank Dr. Fadhlullah, Prof. Maberly, and Prof. Wang for improvement of quality. We also thank Drs. Jing Liu, Jun Zhong, Pan Li, Qiang-Sheng Huang, and Wei-Qi Lu for their help during the sample collection. We thank other members for their help for analysis assistance in Yoshida's laboratory in the Tokyo Institute of Technology, Japan. The authors declare no competing financial interests.

ORCID

Fu-Jun Yue <http://orcid.org/0000-0003-3733-7216> Khan M.G. Mostofa <http://orcid.org/0000-0001-5203-2432> Naohiro Yoshida <http://orcid.org/0000-0003-0454-3849> Shi-Lu Wang <http://orcid.org/0000-0003-0945-8686>

References

- Baggs EM. [2008.](#page-2-0) A review of stable isotope techniques for N_2O source partitioning in soils: recent progress, remaining challenges and future considerations. Rapid Commun Mass Sp. 22:1664–1672.
- Beusen AHW, Bouwman AF, Van Beek LPH, Mogollon JM, Middelburg JJ. [2016](#page-1-0). Global riverine N and P transport to ocean increased during the 20th century despite increased retention along the aquatic continuum. Biogeosciences. 13:2441–2451.
- Casciotti KL, McIlvin M, Buchwald C. [2010](#page-6-0). Oxygen isotopic exchange and fractionation during bacterial ammonia oxidation. Limnol Oceanogr. 55:753–762.
- Finlay JC, Small GE, Sterner RW. [2013](#page-1-0). Human influences on nitrogen removal in lakes. Science. 342:247–250.
- Finlay JC, Sterner RW, Kumar S. [2007.](#page-1-0) Isotopic evidence for in-lake production of accumulating nitrate in Lake Superior. Ecol Appl. 17:2323–2332.
- Frame CH, Casciotti KL. [2010](#page-2-0). Biogeochemical controls and isotopic signatures of nitrous oxide production by a marine

ammonia-oxidizing bacterium. Biogeosciences. 7:2695– 2709.

- Fu PQ, Mostofa KMG, Wu FC, Liu CQ, Li W, Liao HQ, Wang LY, Wang J, Mei Y. [2010](#page-2-0). Excitation-emission matrix characterization of dissolved organic matter sources in two eutrophic lakes (Southwestern China Plateau). Geochem J. 44:99–112.
- Granger J, Sigman DM, Rohde MM, Maldonado MT, Tortell PD. [2010.](#page-2-0) N and O isotope effects during nitrate assimilation by unicellular prokaryotic and eukaryotic plankton cultures. Geochim Cosmochim Ac. 74:1030–1040.
- Heil J, Wolf B, Bruggemann N, Emmenegger L, Tuzson B, Vereecken H, Mohn J. [2014.](#page-2-0) Site-specific ¹⁵N isotopic signatures of abiotically produced N_2O . Geochim Cosmochim Ac. 139:72–82.
- Koba K, Osaka K, Tobari Y, Toyoda S, Ohte N, Katsuyama M, Suzuki N, Itoh M, Yamagishi H, Kawasaki M, et al. [2009.](#page-7-0) Biogeochemistry of nitrous oxide in groundwater in a forested ecosystem elucidated by nitrous oxide isotopomer measurements. Geochim Cosmochim Ac. 73:3115–3133.
- Liu XL. [2010](#page-2-0). The effects of cascade reservoirs on biogeochemical cycling of nitrogen in river-reservoir system – take Wujiang River and its branch Maotiao River in SW China for example. Guiyang (China): University of Chinese Academy of Sciences.
- Liu XL, Liu CQ, Li SL, Wang FS, Wang BL, Wang ZL. [2011.](#page-2-0) Spatiotemporal variations of nitrous oxide (N_2O) emissions from two reservoirs in SW China. Atmos Environ. 45:5458– 5468.
- Mayer B, Wassenaar LI. [2012.](#page-1-0) Isotopic characterization of nitrate sources and transformations in Lake Winnipeg and its contributing rivers, Manitoba, Canada. J Great Lakes Res. 38 (Suppl 3):135–146.
- McIlvin MR, Casciotti KL. [2011](#page-3-0). Technical updates to the bacterial method for nitrate isotopic analyses. Anal Chem. 83:1850–1856.
- Mostofa KMG, Liu CQ, Vione D, Gao KS, Ogawa H. [2013.](#page-1-0) Sources, factors, mechanisms and possible solutions to pollutants in marine ecosystems. Environ Pollut. 182:461–478.
- Mostofa KMG, Liu CQ, Zhai WD, Minella M, Vione D, Gao KS, Minakata D, Arakaki T, Yoshioka T, Hayakawa K, et al. [2016.](#page-6-0) Reviews and syntheses: ocean acidification and its potential impacts on marine ecosystems. Biogeosciences. 13:1767–1786.
- North RL, Winter JG, Dillon PJ. [2013](#page-1-0). Nutrient indicators of agricultural impacts in the tributaries of a large lake. Inland Waters. 3:221–234.
- Ohte N, Tayasu I, Kohzu A, Yoshimizu C, Osaka K, Makabe A, Koba K, Yoshida N, Nagata T. [2010.](#page-1-0) Spatial distribution of nitrate sources of rivers in the Lake Biwa watershed, Japan: controlling factors revealed by nitrogen and oxygen isotope values. Water Resour Res. 46:W07505.
- Pearson LK, Hendy CH, Hamilton DP, Silvester WB. [2012.](#page-1-0) Nitrogen-15 isotope enrichment in benthic boundary layer gases of a stratified eutrophic iron and manganese rich lake. Aquat Geochem. 18:1–19.
- Santoro AE, Buchwald C, McIlvin MR, Casciotti KL. [2011.](#page-2-0) Isotopic signature of N_2O produced by marine ammoniaoxidizing archaea. Science. 333:1282–1285.
- Sasaki Y, Koba K, Yamamoto M, Makabe A, Ueno Y, Nakagawa M, Toyoda S, Yoshida N, Yoh M. [2011.](#page-1-0) Biogeochemistry of nitrous oxide in Lake Kizaki, Japan,

elucidated by nitrous oxide isotopomer analysis. J Geophys Res-Biogeo. 116(G4):G04030.

- Spoelstra J, Schiff SL, Elgood RJ, Semkin RG, Jeffries DS. [2001](#page-1-0). Tracing the sources of exported nitrate in the Turkey lakes watershed using ${}^{15}N/{}^{14}N$ and ${}^{18}O/{}^{16}O$ isotopic ratios. Ecosystems. 4:536–544.
- Sterner RW. [2011.](#page-1-0) C:N:P stoichiometry in lake superior: freshwater sea as end member. Inland Waters. 1:29–46.
- Sutka RL, Adams GC, Ostrom NE, Ostrom PH. [2008](#page-2-0). Isotopologue fractionation during N_2O production by fungal denitrification. Rapid Commun Mass Sp. 22:3989–3996.
- Sutka RL, Ostrom NE, Ostrom PH, Gandhi H, Breznak JA. [2003.](#page-2-0) Nitrogen isotopomer site preference of N_2O produced by Nitrosomonas europaea and Methylococcus capsulatus bath. Rapid Commun Mass Sp. 17:738–745.
- Sutka RL, Ostrom NE, Ostrom PH, Breznak JA, Gandhi H, Pitt AJ, Li F. [2006.](#page-2-0) Distinguishing nitrous oxide production from nitrification and denitrification on the basis of isotopomer abundances. Appl Environ Microb. 72:638–644.
- Toyoda S, Mutobe H, Yamagishi H, Yoshida N, Tanji Y. [2005](#page-2-0). Fractionation of N_2O isotopomers during production by denitrifier. Soil Biol Biochem. 37:1535–1545.
- Toyoda S, Yoshida N. [1999.](#page-2-0) Determination of nitrogen isotopomers of nitrous oxide on a modified isotope ratio mass spectrometer. Anal Chem. 71:4711–4718.
- Toyoda S, Yoshida N. [2016](#page-3-0). Development of automated preparation system for isotopocule analysis of N_2O in various air samples. Atmos Meas Tech. 9:2093–2101.
- Wang SL, Liu CQ, Wang GJ, Tao FX, Li J, Zhu ZZ, Lv YC. [2004.](#page-2-0) Decmposition of organic matter and N_2O cycling in Lake Baihua, Guizhou, during the late stratification. Quaternary Sci. 24:569–577. Chinese with English abstract.
- Well R, Flessa H, Jaradat F, Toyoda S, Yoshida N. [2005](#page-1-0). Measurement of isotopomer signatures of N_2O in groundwater. J Geophys Res-Biogeo. 110:G02006.
- Well R, Flessa H. [2008](#page-7-0). Isotope fractionation factors of N_2O diffusion. Rapid Commun Mass Sp. 22:2621–2628.
- Well R, Flessa H. 2009. Isotopologue signatures of N_2O produced by denitrification in soils. J Geophys Res-Biogeo. 114:G02020.
- Wenk CB, Frame CH, Koba K, Casciotti KL, Veronesi M, Niemann H, Schubert CJ, Yoshida N, Toyoda S, Makabe A, et al. [2016.](#page-1-0) Differential N_2O dynamics in two oxygendeficient lake basins revealed by stable isotope and isotopomer distributions. Limnol Oceanogr. 61:1735–1749.
- Wenk CB, Zopfi J, Blees J, Veronesi M, Niemann H, Lehmann MF. [2014](#page-1-0). Community N and O isotope fractionation by sulfide-dependent denitrification and anammox in a stratified lacustrine water column. Geochim Cosmochim Ac. 125:551–563.
- Wu D, Koster JR, Cardenas LM, Bruggemann N, Lewicka-Szczebak D, Bol R. [2016](#page-2-0). N₂O source partitioning in soils using ^{15}N site preference values corrected for the N₂O reduction effect. Rapid Commun Mass Sp. 30:620–626.
- Xiao HY, Liu CQ. [2004.](#page-1-0) Discrimination between extraneous nitrogen input and interior nitrogen release in lakes. Sci China Ser D. 47:813–821.
- Yamagishi H, Westley MB, Popp BN, Toyoda S, Yoshida N, Watanabe S, Koba K, Yamanaka Y. [2007](#page-2-0). Role of nitrification and denitrification on the nitrous oxide cycle in the eastern tropical North Pacific and Gulf of California. J Geophys Res-Biogeo. 112:G02015.
- Yoshida N, Morimoto H, Hirano M, Koike I, Matsuo S, Wada E, Saino T, Hattori A. [1989](#page-2-0). Nitrification rates and ¹⁵N abundances of N_2O and NO_3^- in the western North Pacific. Nature. 342:895–897.
- Zou Y, Hirono Y, Yanai Y, Hattori S, Toyoda S, Yoshida N. [2014.](#page-7-0) Isotopomer analysis of nitrous oxide accumulated in soil cultivated with tea (Camellia Sinensis) in Shizuoka, central Japan. Soil Biol Biochem. 77:276–291.