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Effects of typical algae species (Aphanizomenon flosaquae and Microcystis aeruginosa) on photoreduction of Hg²⁺ in water body

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ABSTRACT

Photoreduction characteristics of divalent inorganic mercury (Hg²⁺) in the presence of specific algae species are still not well known. Laboratory experiments were conducted in the present study to identify the effects of different concentrations of living/dead algae species, including Aphanizomenon flosaquae (AF) and Microcystis aeruginosa (MA), on the photoreduction rate of Hg²⁺ under various light conditions. The experimental results showed that percentage reduction of Hg²⁺ was significantly influenced by radiation wavelengths, and dramatically decreased with the presence of algae. The highest percentage reduction of Hg²⁺ was induced by UV-A, followed by UV-B, visible light and dark for both living and dead AF, and the order was dark > UV-A > UV-B > visible light for both living and dead MA. There were two aspects, i.e., energy and attenuation rate of light radiation and excrementitious generated from algae metabolisms, were involved in the processes of Hg^{2+} photoreduction with the presence of algae under different light conditions. The percentage reduction of Hg^{2+} decreased from 15% to 11% when living and dead AF concentrations increased by 10 times (from 10^6 to 10^5 cells/mL), and decreased from 11% to ~9% in the case of living and dead MA increased. Algae can adsorb Hg^{2+} and decrease the concentration of free Hg²⁺, thus inhibiting Hg²⁺ photoreduction, especially under the conditions with high concentrations of algae. No significant differences were found in percentage reduction of Hg²⁺ between living and dead treatments of algae species. The results are of great importance for understanding the role of algae in Hg²⁺ photoreduction.

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Introduction

Mercury (Hg) is a highly toxic heavy metal, which can cause detrimental impacts on human and ecosystem health once it enters into aquatic bodies through industrial waste water discharges and atmospheric deposition (Wright et al., 2018). Environmental Hg pollution and its biogeochemical cycling have attracted much attention since the occurrence of Minamata disease (Liu et al., 2012, 2018). Elemental mercury (Hg⁰) has an approximate atmospheric residence time of

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0.5–1 year (O'Driscoll et al., 2005) and thus can be transported globally, causing Hg pollution even in remote areas (Beckers and Rinklebe, 2017). Hg⁰ released across water–air surface is predominantly stimulated by photoreduction of divalent inorganic mercury (Hg²⁺) (Yin et al., 2011). Therefore, the photoreduction of Hg²⁺ plays an important role in the biogeochemical cycle of Hg.

Hg²⁺ reduced to Hg⁰ under UV (280–400 nm) and visible light (400–700 nm) conditions are first- to second-order kinetic reactions (Carla et al., 2015; Zhang, 2006; Zhu et al., 2018). Many factors, such as light conditions, dissolved organic matter, coexisting ions, and biological processes, can influence the photoreduction of Hg²⁺ in water environments (Vost et al., 2012). Algae is a key factor in the process of Hg²⁺ photoreduction in aquatic systems, especially in eutrophic waters where algae are abundant (Lalonde et al., 2004). There are many biologically active and functionalized groups, such as cellulose, pectin substance, alginic acid ammonium salt, polysaccharide, and polygalactose sulphate, on cell surfaces of algae (Deng et al., 2008, 2009; Zeraatkar et al., 2016). Meanwhile, algae have large surface areas with negatively charged cell surface, enabling them to be excellent absorbent to Hg²⁺, which will reduce reactive mercury species in water column (Schartup et al., 2017; Soerensen et al., 2016). Therefore, if the above hypothesis is correct in reality, the reduction rate of Hg²⁺ may decrease in the presence of algae. To date, the characteristics of photoreduction of Hg²⁺ in the presence of certain algae species are still not well known.

Light radiation has been shown to be an important factor affecting the processes of Hg^{2+} photoreduction (Qureshi et al., 2009; Vost et al., 2012). A wide range of wavelengths including UV-A (365 nm), UV-B (310 nm), and visible light interact with dissolved organic matter (DOM), coexisting ions (Fe³⁺, Cl⁻, NO₃, etc.), and algae, thereby generating hydrogen peroxide, hydroxyl radicals, and singlet oxygen, etc., a process that can significantly affect photoreduction of Hg^{2+} (Vost et al., 2012; Zhang, 2006). Light radiation is needed in the process of photosynthesis and thus is also important for the growth of algae. The reaction characteristics of Hg^{2+} reduction under different wavelengths in the presence of algae are still unknown.

Another question is that whether there are any differences in the effects algae on photoreduction of Hg^{2+} between living and dead algae species. Hg^{2+} can cross cell membrane and enter into algae cells by passive transport. In addition, living algae can excrete small organic molecules, such as alginate, halochlamydosan, polypeptide, etc. The excretions also include some free electrons, i.e., hydrated electron (e_{aq}^{-}), hydrogen peroxide (H_2O_2), singlet oxygen ($^{1}O_2$), and superoxide anion (O_2^{-}) (Deng et al., 2008). These excretions can affect the photoreduction of Hg^{2+} through coordinated interactions with small organic molecules and oxidation with free electrons (Vost et al., 2012). However, dead algae do not have such physiological activity, which may cause potential differences between living and dead algae in their impacts on photoreduction of Hg^{2+} .

To address the knowledge gaps outlined above, laboratory experiments were designed to characterize Hg^{2+} reduction under different wavelength ranges in the presence of living/dead algae species, *Aphanizomenon flosaquae* (AF) and Microcystis aeruginosa (MA), both of which are typical algae species in eutrophic water systems. The effects of adsorption process of Hg^{2+} by algae cells on the photoreduction of Hg^{2+} were also determined.

1. Materials and methods

1.1. Experimental materials and devices

A 500-mL quartz glass gas cylinder (diameter 7.2 cm, height 15 cm) was used as the reactor (Fig. 1). The Teflon tubing (Savillex, US) was used as the conduit. Trace mercury was removed by a gold trap amalgam before the carrier gas entering into the reactor. Hg⁰ generated by the Hg²⁺ reduction reaction was separated by high-purity argon (Ar), dried by soda lime (Sigma-Alorich, Germany), and then concentrated onto a gold-coated sand trap. The trapped Hg⁰ was thermally desorbed from the gold sample trap into an inert gas stream (Ar) that carried the released Hg⁰ in the cell of a cold-vapor atomic fluorescence spectrometry (CVAFS) for detection (Model III, Brooks Rand Labs, USA) (EPA, 2002; Sun et al., 2014). UV-A (365 nm, 30 W), UV-B (310 nm, 30 W) and Xenon (visible light, 30 W) lamps were used as light sources. The lamps were 45.5 cm away from the bottle wall. All the bottles were wrapped in tin foil and placed in a dark environment for dark treatment. The temperature of all solutions was 25°C and all experiments were conducted under ambient temperature of 25 ± 0.5°C.

The algal species tested in the present study were AF (Fig. 2A) and MA (Fig. 2B), which were cultured in BG-11 liquid culture medium at 25°C, with illumination intensity of 4000 Lux and the light/dark ratio of 12:12 hr. AF was supposed to be cylindrical with average length and diameter being measured as 9 and 2.23 μ m, respectively. MA was supposed to be spherical with average diameter being measured as 3.42 μ m. Cell surface area to volume ratio was evaluated to be 1.58 and 1.75 μ m²/ μ m³, respectively, for AF and MA.

1.2. Experimental design and methods

The algal samples in stable growth stage were washed and filtrated repeatedly, and then divided into two groups. Algal samples in one group were killed but the cell walls remained intact by water bath calefaction at 50°C for 10 min. The algae concentrations of both groups were diluted to several levels, including 1.0×10^6 , 2.0×10^6 , 4.0×10^6 , 8.0×10^6 , and 10.0×10^6 cells/mL. The cell walls of two algae species kept integrity during a 1440 min period by microscopic observations.

200 mL living/dead algae solution and 20 μ L mercury chloride (HgCl₂, 1000 ng/mL, GR) solution were mixed into the reactor, which was then quickly connected to the pipes as shown in Fig. 1. Light source was turned on simultaneously when high purity argon was added into reaction solution (0.10 L/min). The analytical gold-coated sand trap was replaced at time phases of 20, 60, 120, 240, and 1440 min for measurements. The experimental design is presented in Table 1.



Fig. 1 - Schematic diagram of the reaction device.

Adsorption abilities of algae for Hg^{2+} were also characterized. 200 mL living/dead algae solution (4.0 × 10⁶ cells/mL) and 20 µL mercury chloride (HgCl₂, 1000 ng/mL, GR) solution were added into quartz triangular flask, which was then shaken at a constant speed (120 r/min) under dark condition at the temperature of 25°C. 10 mL solution was sampled from the reactor at every time phase of 20, 60, 120, 240, and 1440 min and then transferred into 10 mL centrifuge tubes (Yonggong, Jiangsu, China). Supernatants were extracted by centrifugation at 4000 r/min for 5 min. The concentrations of total Hg^{2+} in supernatants were determined following the procedures presented in the method 1631 (EPA, 2002). The blank control experiments were conducted using ultrapure water with no algae in the solution. Initial pH values were measured to be 6.3–6.7 for all the experiments.

1.3. Quality control and data analysis

All glass instruments were immersed in 25% nitric acid for 24 hr and then burned in a muffle furnace at 500°C for more than 4 hr before being used, and they were used for only once after naturally cool down in a Hg-free environment. Disposable gloves were worn to prevent cross-contamination. The processes of quality control followed the method 1631 (EPA, 2002). The percentage reduction of Hg²⁺ (%) was calculated as the ratio of total mass of Hg⁰ (ng)/initial amount of Hg²⁺ (ng).

The reaction order was calculated according to the first- and second-order reaction formula (Lu et al., 2004). All the data were analyzed with SPSS 22.0 software for Windows.

2. Results

2.1. Effects of wavelength

The total mass of Hg^0 produced during the 1440 min period under various light conditions ranged from 2.05 to 2.92 ng in the presence of AF, and from 1.81 ng to 2.76 ng in the presence of MA. The order of the produced total Hg^0 mass was UV-A > UV-B > visible light > dark for both living and dead AF, and was dark > UV-A > UV-B > visible light for both living and dead MA (Fig. 3A and B). For blank control, the highest Hg^0 yield was induced by UV-B radiation (5.35 ng), followed by UV-A (4.57 ng), visible light (4.29 ng), and dark (4.08 ng) (Fig. 3C). Variance analysis showed significant differences in the total mass of the produced Hg^0 between the different treatments of algae under various light conditions (p < 0.05).

The Hg⁰ production rates within the first 20 min experiments in the presence of algae were calculated to be (6.35–8.46) × 10^{-4} ng/min under dark, (6.73–9.13) × 10^{-4} ng/min under UV-A, (5.16–8.71) × 10^{-4} ng/min under UV-B, and (5.16–7.57) × 10^{-4} ng/min under visible light. Higher Hg⁰ production



Table 1 – Experiments conducted.		
Objectives	Concentrations of algae species (cells/mL)	Wavelength
Identify the effects of wavelength Identify the effects of algae concentrations Blank control	1.0×10^{6} $1.0 \times 10^{6}, 2.0 \times 10^{6}, 4.0 \times 10^{6}, 8.0 \times 10^{6}, and 10.0 \times 10^{6}$ 0	UV-A, UV-B, visible light, and dark UV-A UV-A, UV-B, visible light, and dark

rates by 1.2–2.8 folds were found for blank control experiments than with algae treatments under various light conditions in this time frame. The Hg⁰ production rate within 240–1440 min under various light conditions were calculated to be $(3.40-3.81) \times 10^{-3}$ ng/min for blank control and $(1.51-2.43) \times 10^{-3}$ ng/min for algae treatments. These results suggested that Hg⁰ production rates were significantly influenced by radiation wavelength, and dramatically decreased with the presence of algae. Hg⁰ production rates were higher in the initial stage than later times during the experiment period.

The percentage reduction of Hg^{2+} under different light conditions was in the decreasing order of UV-A (14.60%) > UV-B (14.06%) > visible light (13.68%) > dark (10.26%) with living AF, dark (13.78%) > UV-A (10.97%) > UV-B (10.73%) > visible light (9.07%) with living MA, UV-A (14.61%) > UV-B (14.26%) > visible light (13.30%) > dark (10.68%) with dead AF, and dark (13.72%) > UV-A (11.40%) > UV-B (10.92%) > visible light (9.30%) with dead MA (Fig. 4). Thus, the percentage reduction of Hg^{2+} varied under different wavelengths regardless of with (or without) living or dead algae. For blank control experiment, the highest percentage reduction of Hg^{2+} was induced by UV-B (26.75%), followed by UV-A (22.86%), visible light (21.45%), and dark (20.41%) (Fig. 4). Comparing results of blank control experiment with those with algae treatments, it was found that the presence of algae inhibited Hg^{2+} photoreduction. Variance analysis elucidated significant differences in the percentage reduction of Hg^{2+} between various light conditions (P < 0.05), indicating wavelength was an important factor impacting the photoreduction of Hg^{2+} with or without algae.

2.2. Effects of algae concentrations

When the AF concentrations were given as 0, 1.0×10^6 , 2.0×10^6 , 4.0×10^6 , 8.0×10^6 , and 10.0×10^6 cells/mL, the respective total mass of Hg⁰ produced were calculated to be 4.57, 2.92, 2.65, 2.59, 2.33, and 2.20 ng for living AF treatments, and were 4.57, 2.92, 2.69, 2.53, 2.37, and 2.25 ng for dead AF treatments (Fig. 5 A). The same tendencies as listed above were also found for the cases with living or dead MA, i.e., total mass of Hg⁰ produced decreased with increasing algae concentrations. Note that there were no significant differences in the total mass of produced Hg⁰ between the cases of living and dead algae treatments (p > 0.05). The total mass of Hg⁰ produced with treatments of living AF, dead AF, living MA, and dead MA decreased by 52%, 51%, 62%, and 61%, respectively, compared with that from the blank control during the 1440 min period (Fig. 5). Correlation analysis showed that total mass of Hg⁰



Fig. 3 – Characteristics of the total mass of Hg⁰ produced under different illumination conditions during 0–1440 min period in the presence of algae.



Fig. 4 – Percentage reduction of Hg²⁺ under different wavelengths of light radiation during the period of 0–1440 min.

produced were negatively correlated to algae concentrations, suggesting that algae species played a key role in controlling the photoreduction of Hg^{2+} , especially at high concentrations. Hg^{0} production rates within 240 min period were calculated to be in the range of (5.75–9.53) × 10^{-3} ng/min from all the algae treatments experiments, which were much lower than that from the



Fig. 5 – Characteristics of Hg⁰ production with the presence of various algae concentrations under UV-A radiation during the period of 0–1440 min.



Fig. 6 – Percentage reduction of Hg²⁺ with the presence of various algae concentrations under UV-A radiation during the period of 0–1440 min.

blank control experiment (18.01 \times 10⁻³ ng/min). The production rates decreased significantly during the rest of the experiment periods ((0.21–0.55) \times 10⁻³ ng/min) for all the treatments.

A significant decreasing tendency was seen in the percentage reduction of Hg^{2+} with increasing algae concentrations (Fig. 6). The percentage reduction of Hg^{2+} decreased from 15% to 11% when living and dead AF concentrations increased by one order of magnitude from 1.0×10^6 to 1.0×10^7 cells/mL, and decreased from 11% to 9% in the case of living and dead MA increased. These results indicated that small differences in Hg^{2+} photoreduction reaction were caused by different algae species. The peak value (22.86%) of percentage reduction of Hg^{2+} was found in the case without algae in reaction solution (Fig. 6).

3. Discussion

3.1. Effects of wavelength

Solar radiation intensity and its wavelength are of great importance for mercury photoreduction in water system (Vost et al., 2012). Photochemical reactions can be either primary or secondary in nature (Zhang, 2006). Ultraviolet radiation is important in aquatic chemistry because of its ability to efficiently induce primary and secondary photochemical reactions in aquatic ecosystems (Häder et al., 2007). Under natural UV light, Hg²⁺, such as Hg(OH)₂, HgSO₃, and Hg(HSO₃)⁻, can all undergo reduction reactions to generate dissolved gaseous mercury (DGM) in natural water systems (Sun et al., 2015). Ultraviolet light promotes the production of DGM (Sun et al., 2015), of which 61%-73% were produced under UV light, and a maximum of 27% was produced under visible light (Garcia et al., 2005). Initial work suggested that UV-A radiation was primarily responsible for mercury reductive reactions because DGM production rates did not change significantly by



Fig. 7 – Concentrations of Hg²⁺ in supernatants for adsorption experiment during the period of 0–1440 min.

the removal of UV-B radiation using a Mylar screen (Amyot et al., 1994). A recent study suggested that UV-B radiation was a key portion of electromagnetic spectrum responsible for mercury reduction in freshwaters (Vost et al., 2012). The controlled radiation experiments in freshwaters found that Hg^{2+} photoreduction rate constant was higher for UV-B radiation than for UV-A radiation (O'driscoll et al., 2006a; O'Driscoll et al., 2006b). At present, the exact mechanism of UV light-induced Hg^{2+} photoreduction is still unclear.

Indeed, Hg²⁺ photoreduction rate is affected by a combination of the wavelength and intensity of radiation, the chemical composition of water body, the concentration and structure of chromophores in the natural water, and the rate of attenuation of specific radiation wavebands (Vost et al., 2012). UV-B radiation is shorter in wavelength and higher in energy than UV-A radiation (Vost et al., 2012). The 6s orbital of mercury is assumed to be the acceptor orbital in all wavelength ranges, resulting in different excited states generated via transitions from the steady state by absorbing different spectrum (Sun et al., 2014; Zhang, 2006). Therefore, we found that the highest percentage reduction of Hg²⁺ was induced by UV-B radiation, followed by UV-A radiation, visible light and dark in the absence of algae species. However, the presence of algae species disturbed this order. Incident radiation of UV-B radiation was lower than that of UV-A radiation because of the more-rapid attenuation of UV-B. The reaction rates with the presence of AF were thus higher under UV-A radiation than UV-B radiation. The energies of quantum photon of visible light were too low to induce photoreduction of Hg²⁺, in contrast to the case found under UV radiations. Under dark conditions, there were no quantum photons to excite photochemical reactions. Therefore, the percentage reduction of Hg²⁺ with the presence of AF were in the decreasing order of UV-A > UV-B > visible light > dark. As for the cases with the presence of MA, some other processes affected photoreduction of Hg²⁺. Highly reactive radicals, i.e., hydrated electrons (e_{aq}) , superoxide anion (O_2) , hydrogen peroxide (H_2O_2) , and singlet oxygen (¹O₂), which were generated from some algae metabolisms Eqs. (1)–(4), can inhibit the process of Hg^{2+}



Fig. 8 – Adsorption quantity of Hg^{2+} by AF and MA during the period of 0–1440 min.

reduced to Hg^0 under light conditions. Thus, the highest percentage reduction of Hg^{2+} was found under dark condition with the presence of MA. Some other factors, such as metabolisms of algae and mercury–algae complex reactions, or some other mechanisms may also be involved in Hg^{2+} photoreduction, resulting in very complex processes with the presence of algae species, especially in natural water systems. Investigating the roles of specific algae in the photoreduction processes of Hg^{2+} is of great importance for understanding biogeochemical and cycling characteristics of mercury.

$$\mathbf{e}_{aq}^{-} + \mathbf{O}_{2} \rightarrow \mathbf{O}_{2}^{-} \tag{1}$$

$$2O_2^- + 2H^+ \rightarrow H_2O_2 + O_2$$
 (2)

$$\operatorname{Chl} + hv \longrightarrow^{1} \operatorname{Chl}^{2} \operatorname{Chl}$$
 (3)

$$1Chl + O_2 \rightarrow 1O_2 + Chl \tag{4}$$

3.2. Effects of adsorption

The concentrations of Hg²⁺ in supernatants decreased significantly in all the treatments with the presence of living and dead algae, but did not change in the blank control experiment during the 1440 min experiment period (Fig. 7). This demonstrates that adsorption process played a key role in decreasing the concentrations of free Hg^{2+} in water. The adsorption quantity of Hg²⁺ by the algae cells rapidly increased during the first 5 min. The adsorption quantity was 1.45 ng higher from living than dead AF (7.22 vs. 5.77 ng), and 0.22 ng higher for the case of MA (8.45 vs. 8.23 ng) (Fig. 8). The changes in the adsorption amounts slowed down during 5-120 min and tended to stabilize during 120-1440 min. The respective concentrations of Hg2+ decreased to 56.31 and 50.16 ng/L at 1440 min in the presence of living AF and MA, and to 64.67 and 52.38 ng/L in the presence of dead AF and MA. This showed that the highest adsorbing capacity of adsorbent was living MA, followed by dead MA, living AF, and dead AF. Such differences in the adsorbing capacity between the two algae species were due to the higher cell surface area to volume ratio of MA than AF, and thus greater capacity for adsorbing Hg²⁺.

The adsorption of Hg^{2+} by algae cells mainly consists of passive extracellular adsorption and active accumulation. Passive adsorption is the predominant process with a

relatively fast rate (Diéguez et al., 2013). Hg²⁺ absorbed on cell surface of dead algae is the same as the process of passive adsorption by living algae (Zeraatkar et al., 2016). The living algae can adsorb Hg²⁺ through active accumulation as well as passive extracellular adsorption. Both adsorption processes can decrease the concentrations of free Hg²⁺, thus declining the percentages of reduced Hg2+. Although the adsorption processes are somewhat different between living and dead algae, no significant differences were found in the percentage reduction of Hg²⁺ in our experiments, indicating that after the addition of $HgCl_2$, the adsorption of Hg^{2+} by living and dead algae cells were mainly passive adsorption. This is because the process of active accumulation is very slow and plays a negligible role in the reactions. Algae may affect the concentration and structure of available DOM, thereby changing the chemical conditions of the photoreaction (Vost et al., 2012). If biological process is the major factor in affecting Hg²⁺ photoreduction, the reaction rates should change slowly in the initial stage and then change quickly at later stages because metabolism of algae could further change the chemical conditions. Photobiological reduction by algae may be mainly controlled by the excretion of small organic molecules and free electrons (Vost et al., 2012). Previous studies have identified no difference in DGM production between the presence of either diatom and its isolated exudates (Deng et al., 2008; Lanzillotta et al., 2004). Thus, physical adsorption is believed to be the predominant process after adding HgCl₂.

The cell wall of algae contains polysaccharides, proteins, amino, carbonyl, carboxyl, sulfhydryl, etc. groups that can bind to Hg²⁺ and adsorb Hg²⁺ to the cell wall (Schartup et al., 2017). Deng et al. (2008) found that the photoreduction rate of Hg²⁺ increased with the increasing concentration of Anabeana, and an opposite trend was found in the present study, likely due to the different cell structures of different algae species used here, which have different adsorption effect on Hg²⁺ photoreduction. AF and MA belong to Cyanophyta, which has a good adsorption effect on heavy metals (Zeraatkar et al., 2016), and thus has a certain influence on the photochemical reaction of Hg²⁺. pH value may be an important factor affecting absorption because of its influence on surface charge and cation exchange capacity. However, pH value was around 6.5 and did not change much during the whole 1440 min experiment period, preventing us from conducting further indepth analysis on this issue. Whether DGM can be adsorbed on algae surface or not remain unclear in our experiment. Future research should focus on identifying species and concentrations of Hg in algae cell and estimating the role of algae species in biogeochemistry of Hg in water environment system.

4. Conclusions

Light radiation had an important influence on the reduction of Hg²⁺, and the effects of different light wavelengths on the reduction process of Hg²⁺ were quite different. The photoreduction of Hg²⁺ was inhibited by *Aphanizomenon flosaquae* (AF) and *Microcystis aeruginosa* (MA) with more obvious inhibitory effects identified under high algae concentrations. The percentage reduction of Hg^{2+} was higher in the presence of AF than MA, suggesting algae species being an important factor in Hg^{2+} photoreduction. There were no significant differences between living and dead algae treatments as indicated by their similar reaction rates. Algae can adsorb Hg^{2+} and thus decrease the concentrations of free Hg^{2+} in solution, however, it is not clear whether the produced Hg^{0} was generated from the photoreduction of free Hg^{2+} in solution or from algae–Hg complexes. The biogeochemistry of algae–Hg complexes should be explored further.

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