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Phytolith carbon sequestration in bamboos of different ecotypes: a case study in China

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Abstract Occlusion of carbon (C) within phytoliths (PhytOC) is becoming one of the most promising terrestrial C sequestration mechanisms. This study explored the production of PhytOC within 35 bamboo species belonging to three ecotypes using methods of microwave digestion. The aim of this study is to explore the present and potential C sequestration rate within phytoliths of bamboo species from three ecotypes. PhytOC content in bamboos of three ecotypes ranges from 0.07 % to 0.42 %. The mean PhytOC production flux decreases as: clustered bamboo (0.050 \pm $0.016 \text{ t CO}_2 \text{ ha}^{-1} \text{ a}^{-1}) \approx \text{ mixed bamboo } (0.049 \pm 0.016 \text{ t})$ $CO_2 ha^{-1} a^{-1}$ > scattered bamboo (0.038 ± 0.020 t CO_2 $ha^{-1}a^{-1}$). The phytolith carbon sequestration in Chinese bamboo is estimated to be 0.293 ± 0.127 Tg (1 Tg = 10^{12} g) CO₂ a⁻¹; approximately 75 %, 3 %, and 22 % of which is contributed from scattered, mixed and clustered bamboo, respectively. Taking the PhytOC production flux of 0.18 ± 0.12 t CO₂ ha⁻¹ a⁻¹ and current annual area increasing rate of 3 %, global bamboo phytoliths would

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sequester 11.9 \pm 7.9 Tg CO₂ a⁻¹ by 2050. Consequently, bamboo forests have significant potential to mitigate the increasing concentration of atmospheric CO₂ by maximizing PhytOC production flux and expanding bamboos.

Keywords Bamboo · Carbon sequestration · Ecotypes · Phytolith · PhytOC

1 Introduction

Global climate change is one of the adverse effects of the increasing concentration of atmospheric greenhouse gas [1-3]. At present, the global concentration of atmospheric CO₂ has reached about 391 parts per million by volume [4] and the rate of increase is expected to be significant in the future due to the growing anthropogenic and natural emissions of CO₂, such as emissions from transportation, industry, electrical generation and building operations [1]. Removing atmospheric C and sequestrating it in the terrestrial biosphere is considered to be an effective way of reducing levels of atmospheric CO₂ [5–8].

The occlusion of carbon within phytoliths is one of the most promising mechanisms of biotic carbon sequestration [7, 8]. PhytOC is deposited within plants tissues (mainly in leaves) and released into the soil after plant decomposition [9–11]. PhytOC is a relatively stable form of C and has been demonstrated to represent up to 82 % of soil carbon in some sediments after 2000 years because of its strong ability to resist decomposition [10, 12–14].

Grasses and herbaceous plants such as bamboo, sugarcane, silvergrass and bulrush have been demonstrated to have considerable capacities to sequestrate CO_2 during the formation of phytoliths [2, 3, 7, 8, 11, 15]. The PhytOC production fluxes in economic bamboo [11], sugarcane [15], wheat [16] and millet [17] have been reported to be 0.7, 0.36, 0.246 and 0.03 t CO₂ ha⁻¹ a⁻¹, respectively. These results suggest that bamboo has a larger potential of phytolith carbon sequestraion than other plant species.

Forest ecosystems sequester up to 80 % of total aboveground and 40 % of below-ground C of the earth, thus playing an important role in the global C balance [18–21]. Relative to other forests such as Chinese fir and Masson pine [22], bamboo forests have a stronger ability to sequester carbon because of their faster growth rate and easier propagation and regeneration [11, 23]. Bamboo forests are mainly distributed in the subtropics and tropics of South and Southeast Asia (particularly in China), Africa, and Latin America [6, 24] with a global area of 22×10^6 ha, accounting for about 1 % of the total area of the world's forests [22, 25]. Although the area of world's forests has decreased drastically, the bamboo forest area has increased at an annual rate of 3 % [25].

The area of bamboo in China is approximately 7.2×10^6 ha and is increasing progressively [6, 22]. During the past 30 years in China, afforestation and reforestation of bamboo and other forests have been successful in mitigating the ever increasing atmospheric CO₂ concentration by capturing it in living biomass [26, 27]. Although Parr et al. [11] had reported the phytolith carbon sequestration of bamboo, their study was focused on too limited bamboo species (10 species) and the factors of bamboo ecotype differences were not taken into full consideration. This study investigated the rates of phytolith production and phytolith carbon sequestration in 35 bamboo species of three different ecotypes in China. The purposes of this study are to examine the present and potential phytolith carbon sequestration rate of bamboo and offer references for the regulation of atmospheric CO₂ through phytolith C sequestration.

2 Materials and methods

2.1 Collection of bamboo leaves and soils

Based on morphological characteristics of underground stem, bamboos are usually divided into three ecotypes including clustered, scattered and mixed bamboos [20, 22, 28]. Clustered, scattered and mixed bamboos are originally distributed in tropical, subtropical, and relatively low temperature areas of the world, respectively [20, 22, 28]. The 35 dominant bamboo species of three ecotypes were sampled from a bamboo garden at Zhejiang Agricultural and Forestry University (30°15'N, 119°43'E), Lin'an, Zhejiang, China. The bamboo garden was built in 2000 and most of its bamboo species originate from southern China and Japan. Lin'an has a subtropical monsoonal climate. The frost-free period lasts up to 234 days, with an average of 1,400 mm rainfall annually, and the area has an annual average temperature of 16 °C. The soil of the site is acidic with a mean pH of 5.47. The contents of soil organic matter and total SiO₂ were 2.61 % and 57.81 %, respectively. The available Si content was 195.49 mg kg⁻¹.

The accumulation of Si is known to be greater in mature plant organs than in other organs [29–31]; we thus selected mature bamboo leaves (two year old) to ensure the maximum accumulation of Si. The mixing leaves from up to down of 35 bamboo species (three replicates) were sampled with a special sickle. Each bamboo sample made up about 300 g of composite mature leaf litters. Samples were rinsed with ultrapure water three times, oven-dried at 75 °C for 48 h, and cut into small pieces for chemical analysis.

Surface soil samples were collected near the base of bamboo, air-dried and divided into two subsamples. One subsample was passed through 20 mesh sieve and used to analyze soil pH. The other subsample was passed through 100 mesh sieve after removing root residua, and used to analyze contents of SOM, total SiO₂ and available Si [32].

2.2 Analysis of Si, phytoliths and PhytOC

After ashing and Li-metaborate fusion, samples were dissolved in dilute nitric acid and analyzed by spectrophotometry for Si content [32]. The bamboo phytoliths were extracted using a microwave digestion procedure [33]. Possible extraneous organic material of phytoliths was examined and removed with $0.8 \text{ mol } L^{-1}$ potassium dichromate [2, 3, 11, 34]. The cleaned phytolith samples were oven-dried at 70 °C for 24 h in a centrifuge tube, and weighed to obtain the content of phytoliths. All phytolith samples were checked with an optical microscope (Olympus CX31, Japan) to ensure that all possible extraneous organic material was removed [35]. The phytoliths were then treated with 4 mol L^{-1} hydrogen fluoride (HF) at 45 °C for 60 min to dissolve phytolith-Si [36]. The organic C content within phytoliths after HF treatment was determined by the potassium dichromate method [32]. The PhytOC determination was checked with standard samples of GBW07405. The precision was better than 7 %.

2.3 Estimation of PhytOC production

The PhytOC production flux of bamboo leaf litter can be estimated from the PhytOC content of biomass and ANPP of leaf litter as

PhytOC production flux = PhytOC content $\times 44/12$ \times ANPP of leaf litter, (1)

where PhytOC production flux is PhytOC production amount of bamboo leaf litter per area per year (kg CO_2 $ha^{-1}a^{-1}$), PhytOC content is the PhytOC content in bamboo leaf litter biomass (wt%), and ANPP of leaf litter is aboveground net primary productivity of bamboo leaf litter (kg $ha^{-1}a^{-1}$).

PhytOC production rate of bamboo leaf litter can be estimated from data of PhytOC flux and bamboo area as

PhytOC production rate = PhytOC production flux

$$\times$$
 area, (2)

where PhytOC production rate is total PhytOC production amount in bamboo leaf litter per year (kg $CO_2 a^{-1}$), PhytOC production flux is estimated from Eq. (1), area is the area of bamboo (ha) [7].

Student's *t*-test and Excel and SPSS software were employed in the statistical analysis of all data.

3 Results

The dry weight of SiO₂, phytolith and PhytOC contents among the 35 bamboo species were 5.41 %–17.62 %, 3.47 %–16.28 % and 0.07 %–0.42 %, respectively (Table 1). However, the average contents of SiO₂ (9.22 %– 13.35 %), phytoliths (7.69 %–11.09 %) and PhytOC (0.17 %–0.21 %) in leaf litter decreased in the following order: clustered \approx mixed > scattered bamboo.

A significant correlation existed between the content of SiO₂ and phytoliths within the 35 bamboo species ($R^2 = 0.89$, P < 0.01) (Fig. 1). Phytolith content in leaf litter and PhytOC content in phytoliths were non-significant ($R^2 = 0.0255$, P > 0.05) (Fig. 2a). The correlation between PhytOC content in leaf litter and phytolith content of biomass was significant ($R^2 = 0.4588$, P < 0.01) (Fig. 2b). A significant correlation ($R^2 = 0.3442$, P < 0.05) (Fig. 2c) existed between PhytOC content in leaf litter and PhytOC content in phytoliths.

4 Discussion

4.1 Mechanisms of carbon sequestration within phytoliths of bamboo

The significantly possitive correlation among contents of SiO₂ and phytoliths in leaf litter of 35 different bamboo species ($R^2 = 0.89$, P < 0.01) (Fig. 1) demonstrate that the decreasing trend of phytolith contents in leaf litter (clustered \approx mixed > scattered bamboo) (Table 1) can be attributed to different absorption capacities of Si in three ecotypes of bamboos. Ding et al. [37] and Parr et al. [11] demonstrated that the abilities of Si absorption varied among bamboo species possibility due to their different growth characteristics and habits, with stronger Si

accumulation capacities in clustered and mixed bamboo than in scattered bamboo [11, 37] (Table 1). However, the exact mechanism and controlling factors of Si uptake difference among different bamboo species and ecotypes require further study.

In previous studies, many researchers found that dry weight of PhytOC in sugarcane, economic bamboo species, wheat and millet had no correlation with silica content [11. 15-17]. Moreover, they reported that the amount of C sequestered in plants mainly depended on the plant's C sequestration capacity during the formation of phytoliths rather than the actual silica taken up by plants. However, the results of this study are quite different. The significant correlations between the PhytOC content in leaf litter and phytolith content ($R^2 = 0.4588$, P < 0.01) (Fig. 2b) and between the PhytOC content in leaf litter and PhytOC content in phytoliths ($R^2 = 0.3442$, P < 0.05) (Fig. 2c) in this study indicate that the final production of PhytOC depends not only on the efficiency of phytolith carbon sequestration, but also on the actual phytolith production rate during plant growth. Thus, all possible measures that can improve the Si content of plants and the capacity of C occlusion in the phytoliths could be taken to increase the content of PhytOC in leaf litter. The applications of Si fertilizers [38–41] and Si-rich organic mulches [42, 43] have been shown to play an important role in enhancing the Si content and PhytOC accumulation during plant growth. In addition, applying fertilizers to increase the ANPP of bamboo may also increase the PhytOC production flux. Besides the increasing of PhytOC production flux, the PhytOC production rate can also be improved through the expanding distribution area of bamboo.

4.2 The phytolith carbon sequestration in Chinese bamboos

The leaf litter production flux in clustered, mixed and scattered bamboo in China is reported to be 5.59–7.04 t ha⁻¹ a⁻¹, 3.77–8.14 t ha⁻¹ a⁻¹ and 5.90–6.11 t ha⁻¹ a⁻¹, respectively [44–48]. The PhytOC production flux in bamboo can be estimated using Eq. (1). The PhytOC flux in studied bamboo varies from 0.017 to 0.081 t CO₂ ha⁻¹ a⁻¹, with a mean of 0.041 t CO₂ ha⁻¹ a⁻¹. The flux among the three ecotypes of bamboo decreases in the order: clustered bamboo \approx mixed bamboo > scattered bamboo (Table 2).

Bamboo distribution area in China is 7.2×10^6 ha [22]. The area of scattered, mixed and clustered bamboo is 5.76×10^6 ha, 0.14×10^6 ha and 1.30×10^6 ha, respectively [20, 49, 50]. The PhytOC production rate in bamboo can be estimated using Eq. (2). Based on these data and taking the mean PhytOC production flux of three ecotypes of bamboo, the current rate of C sequestration within phytoliths (PhytOC) in Chinese bamboo is

Table 1 Contents of SiO₂, phytoliths and PhytOC among 35 bamboo species of different ecotypes (%)

Ecotypes	Bamboo species	SiO_2 in leaves		Phytoliths in leaves		PhytOC in leaves	
			SD	Mean	SD	Mean	SD
Scattered bamboo	Chimonobambusa quadrangularis	5.41	1.01	5.16	0.20	0.22	0.012
	Phyllostachys vivax McClure cv. aureocaulis	9.21	0.76	8.17	0.21	0.12	0.010
	Phyllostachys. parvifolia	9.84	0.68	8.58	0.57	0.19	0.019
	Phyllostachys imcarnata	9.53	1.10	8.75	0.05	0.18	0.009
	Phyllostachys vivax f. huanwenzhu J.L. Lu	8.33	1.07	6.83	0.12	0.16	0.014
	Phyllostachys aureosulcata	11.25	1.24	9.56	0.13	0.20	0.016
	Phyllostachys bambusoides var.castillonis	10.47	0.87	9.28	0.25	0.17	0.012
	Phyllostachys bambusoides	12.75	0.81	11.20	0.22	0.18	0.003
	Phyllostachys aureosuleata McClure cv. Pekinensis J.L. Lu	8.74	0.21	6.72	0.59	0.09	0.013
	Phyllostachys. heterocycla cv. huamozhu	9.14	1.27	6.87	0.23	0.07	0.009
	Phyllostachys propinqua	12.64	1.24	11.10	0.40	0.42	0.009
	Phyllostachys heterocycla (Carr.) Mit ford	8.63	1.52	5.18	0.26	0.11	0.008
	Phyllostachys prominens	5.51	0.86	3.47	0.33	0.10	0.006
	Phyllostachys. Arcane	11.20	0.14	9.42	1.17	0.27	0.092
	Phyllostachys pubescens	5.61	1.18	5.08	1.03	0.09	0.027
	Mean	9.22	0.93	7.69	0.38	0.17	0.017
Mixed bamboo	Sinobambusa tootsik	14.62	1.69	13.45	0.62	0.28	0.076
	Hibanobambus tranguillans	11.40	0.63	9.75	0.27	0.16	0.005
	Shibataea chinensis Nakai	9.51	0.87	8.53	0.23	0.27	0.005
	Shibataea chinensis	9.61	1.76	8.60	0.54	0.11	0.014
	Shibataea kumasasa	11.86	2.62	9.50	0.28	0.19	0.015
	Acidosasa gigantea	9.84	1.11	6.59	0.43	0.12	0.011
	Oligostachyum sulcatum	11.83	0.72	8.59	0.50	0.13	0.016
	Pseudosasa japonica	11.31	0.10	8.87	0.09	0.13	0.040
	Pseudosasa japonica var. Tsutsumiana Yanagita	11.81	0.34	9.00	0.12	0.16	0.025
	Pleioblastus kongosanensis	17.62	1.40	12.38	0.29	0.29	0.019
	Sasa auricoma	17.17	2.43	16.28	0.37	0.31	0.031
	Sasa argenteastriatus	12.37	0.47	10.33	0.15	0.23	0.007
	Mean	12.41	1.18	10.16	0.32	0.20	0.022
Clustered bamboo	Bambusa multiplex f. alphonsekarri	12.33	1.15	11.07	0.27	0.24	0.022
	Bambusa multiplex cv. Fernleaf	14.52	2.80	12.39	0.25	0.16	0.010
	Bambusa multiplex (Lour.) RaeuschelexJ.A.etJ.H.Schul.	10.18	1.73	8.17	0.26	0.15	0.013
	Dendrocalamus latifloru	17.06	0.57	13.69	0.41	0.28	0.020
	Bambusa vulgaris	12.49	1.26	11.30	0.15	0.15	0.020
	Bambusa tuldoides	14.72	0.16	11.76	0.14	0.24	0.030
	Dendrocalamopsis oldhami	15.75	0.18	12.58	0.16	0.26	0.020
	Bambusa multiplex cv. Alphonse-Karr	9.73	0.15	7.78	0.06	0.17	0.020
	Mean	13.35	1.00	11.09	0.21	0.21	0.019

 0.293 ± 0.127 Tg CO₂ a⁻¹; 75 %, 3 % and 22 % of which are contributed from scattered, mixed and clustered bamboo, respectively (Table 2). Due to the different PhytOC production fluxes and planting areas, the PhytOC production rates vary significantly among the three bamboo ecotypes, and the rate of scattered bamboo is significantly higher than that of mixed or clustered bamboo. The content of PhytOC in dry leaf biomass and

the estimated phytolith carbon sequestration fluxes and rates in our study are lower than those found by Parr et al. [11]. This could be explained by (1) the lower ANPP of Chinese bamboo estimated in this study (3.77–8.14 t $ha^{-1} a^{-1}$) as compared to that in Parr et al. [11] and (2) the selection of the mean PhytOC production flux rather than the median value in our PhytOC production rate estimation.



Fig. 1 Correlation of phytolith content and SiO_2 content in bamboo leaf litter

Although the production rate of PhytOC in bamboo is lower than grasslands and fens due to its smaller distribution area, the phytolith C sequestration flux of bamboo is higher than that of grassland, millet and fen (Table 3).

4.3 The implication for phytolith carbon sequestration in global bamboos

Taking the mean PhytOC production flux of 0.041 t CO₂ $ha^{-1}a^{-1}$ (Table 2) and the current global bamboo area of 22×10^6 ha, approximately 0.89 Tg CO₂ from the atmosphere would be sequestrated globally by various bamboo species every year. It is possible to enhance the potential phytolith carbon sequestration rate by improving the actual phytolith production flux of bamboo and increasing the area of bamboo forest. Although the phytolith carbon production flux of bamboo in our study is low, it is possible to improve it to 0.18 ± 0.12 t CO₂ ha⁻¹ a⁻¹ (half of the result of Parr et al. [11]) by regulating silicon supply such as the applications of Si fertilizers [38–41] and Si-rich organic mulches [42, 43]. The current global area of bamboo forest is 22×10^6 ha, however, it will have tripled by 2050 given the current annual area growth rate of 3 % [11, 25]. Taking the production flux $(0.18 \pm 0.12 \text{ t CO}_2 \text{ ha}^{-1} \text{ a}^{-1})$ of PhytOC and the tripled area, the phytoliths of global bamboo would be expected to sequester $11.9 \pm 7.9 \text{ Tg CO}_2 \text{ a}^{-1}$ from the atmosphere in 2050.

In comparison with other plant ecosystems (Table 4), the global potential rate of bamboo PhytOC production in this study and that of Parr et al. [11] is similar to that of rice due to its smaller distribution area [2], but higher than that of sugarcane, fen and millet [3, 15, 17].

Sequestration of atmospheric CO_2 in the terrestrial biosphere is an effective mechanism for reducing atmospheric



Fig. 2 Correlations among C content of phytolith, phytolith content and PhytOC content in leaf litter. **a** C content of phytolith vs phytolith content; **b** PhytOC content in leaf litter and phytolith content; **c** PhytOC content in leaf litter and C content in phytolith

 CO_2 concentration. It is possible to relieve some of the adverse impacts of climate change by selecting and growing plants with high phytolith content and a strong capacity to

 Table 2
 The mean PhytOC production fluxes and rates of three ecotypes of bamboo (SD)

Ecotypes	Area (10 ⁶ ha)	Estimated PhytOC fluxes (t CO_2 ha ⁻¹ a ⁻¹)	Estimated PhytOC rates $(Tg CO_2 a^{-1})^b$
Scattered bamboo	5.76	0.038 (0.020)	0.220 (0.110)
Mixed bamboo	0.14	0.049 (0.016)	0.007 (0.002)
Clustered bamboo	1.30	0.050 (0.016)	0.065 (0.015)
Total	7.20	0.041 (0.017)	0.293 (0.127)
b 1 Tg = 1	10^{12} g		

Table 3 Comparison of estimated PhytOC fluxes and PhytOC production rates in different ecosystems of China

Plant species	PhytOC in leaves (%)	PhytOC production fluxes (t CO_2 ha ⁻¹ a ⁻¹)	PhytOC production rate $(Tg CO_2 a^{-1})$	Ref.
Bamboo	0.07–0.31	0.017-0.081	0.292 (0.120)	This study
Grassland	0.04-0.06	0.001-0.002	0.600 (0.171)	[7]
Millet	0.04-0.27	0.008-0.038	0.042 (0.028)	[17]
Fen	0.01-0.25	0.003-0.077	0.545 (0.493)	[3]

The number within parenthesis is standard deviation

Table 4 Comparison of estimated PhytOC fluxes and global potential PhytOC production rates in different ecosystems

Plant species	PhytOC production fluxes (t CO_2 ha ⁻¹ a ⁻¹)	Potential PhytOC production rates $(Tg CO_2 a^{-1})$	Ref.
Bamboo ^a	0.067–0.319	4.0–19.8	This study
Economic bamboo	0.008-0.709	7.8–15.6	[11]
Sugarcane	0.120-0.360	2.4–7.2	[15]
Rice	0.026-0.125	3.9–19.4	[2]
Fen	0.003-0.077	1.0-11.4	[3]
Millet	0.008-0.038	0.5–2.7	[<mark>17</mark>]

^a The potential area of world's bamboo is expected to reach 66×10^6 ha by 2050 [11, 25]

occlude C within their phytoliths [11]. In spite of some uncertainties in calculating bamboo phytolith and PhytOC production, caused mainly by regional or species variation, the results of the study indicate that bamboo phytoliths have great potential to occlude CO_2 because of their outstanding peculiarities of high phytolith content, rapid growth and reproduction and ease of regeneration. Therefore, bamboo forests have significant potential to mitigate the increasing concentration of atmospheric CO_2 if maximizing the production of PhytOC and expanding bamboo distribution area.

5 Conclusions

The PhytOC content of different bamboo species of three ecotypes (scattered, mixed and clustered bamboo) ranges from 0.07 % to 0.42 %. Generally, the dry mass of phytoliths and PhytOC within the three ecotypes of bamboo decrease in the order: clustered \approx mixed > scattered bamboo. The data indicate that the mean PhytOC production flux likewise decreases in the order: clustered bamboo $(0.050 \pm 0.016 \text{ t CO}_2 \text{ ha}^{-1} \text{ a}^{-1}) \approx \text{ mixed bamboo} (0.049 \pm 1000 \text{ cm}^{-1} \text{ cm}^{-1} \text{ cm}^{-1})$ $0.016 \text{ t CO}_2 \text{ ha}^{-1} \text{ a}^{-1}) > \text{scattered bamboo} (0.038 \pm 0.020 \text{ t})$ CO_2 ha⁻¹ a⁻¹). The C sequestration rate within phytoliths (PhytOC) in Chinese bamboo is estimated to be about 0.293 ± 0.127 Tg CO₂ a⁻¹; 75 %, 3 % and 22 % of which are contributed from scattered, mixed and clustered bamboo, respectively. Taking the potential production flux of PhytOC $(0.18 \pm 0.12 \text{ t CO}_2 \text{ ha}^{-1} \text{ a}^{-1})$ and potential bamboo area $(66 \times 10^6 \text{ ha}), 11.9 \pm 7.9 \text{ Tg CO}_2 \text{ a}^{-1}$ from the atmosphere would be sequestered in bamboo phytoliths globally in 2050.

Although our method is not complex, we think that our study offers a rough estimation of phytolith carbon sequestration in bamboo forest of China and of the world, which allows for direct comparison of phytolith carbon sequestration among different terrestrial ecosystems and offers references for bamboo afforestation or reforestation practices. Further study on the effects of management practices (e.g., rock-powder amendment and silica fertilization) on bamboo phytolith C sequestration is required before practices of bamboo phytolith C sequestration enhancement can be used to reduce the concentration of greenhouse gases in the atmosphere at a regional or global scale.

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Conflict of interest The authors declare that they have no conflict of interest.

References

- Intergovernmental Panel on Climate Change (IPCC) (2007) Climate change 2007: the scientific basis. Cambridge University Press, Cambridge. http://www.slvwd.com/agendas/Full/2007/06-07-07/Item%2010b.pdf
- Li ZM, Song ZL, Parr JF et al (2013) Occluded C in rice phytoliths: implications to biogeochemical carbon sequestration. Plant Soil 370:615–623

- Li ZM, Song ZL, Jiang PK (2013) Biogeochemical sequestration of carbon within phytoliths of wetland plants: a case study of Xixi wetland, China. Chin Sci Bull 58:2480–2487
- 4. Stocker T, Qin D, Plattner G et al (2013) Climate change 2013: the physical science basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge. http:// www.climatechange2013.org/images/report/WG1AR5_Frontmatter_ FINAL.pdf
- Oldenburg CM, Torn MS, DeAngelis KM et al (2008) Biologically enhanced carbon sequestration: research needs and opportunities. Report on the energy biosciences institute workshop on biologically enhanced carbon sequestration. http://www.climatechange 2013.org/images/report/WG1AR5_Frontmatter_FINAL.pdf
- Zhou GM, Zhuang SY, Jiang PK et al (2011) Soil organic carbon accumulation in intensively managed *Phyllostachys praecox* Stands. Bot Rev 77:296–303
- Song ZL, Liu HY, Si Y et al (2013) The production of phytoliths in China's grasslands: implications to the biogeochemical sequestration of atmospheric CO₂. Glob Change Biol 18:3647–3653
- Song ZL, Wang HL, Strong PJ et al (2012) Plant impact on the coupled terrestrial biogeochemical cycles of silicon and carbon: implications for biogeochemical carbon sequestration. Earth Sci Rev 115:319–331
- 9. Piperno DR (1988) Phytolith analysis: an archaeological and geological. Academic Press. Inc., London
- Parr JF, Sullivan LA (2005) Soil carbon sequestration in phytoliths. Soil Biol Biochem 37:117–124
- Parr JF, Sullivan LA, Chen B et al (2010) Carbon bio-sequestration within the phytoliths of economic bamboo species. Glob Change Biol 16:2661–2667
- Wilding LP, Drees LR (1974) Contributions of forest opal and associated crystalline phases to fine silt and clay fractions of soils. Clay Clay Miner 22:295–306
- Mulholland SC, Prior CA (1993) AMS radiocarbon dating of phytoliths. MASCA Res Pap Sci Archaeol 10:21–23
- Prasad V, Strömberg C, Alimohammadian H et al (2005) Dinosaur coprolites and the early evolution of grasses and grazers. Science 310:1177–1180
- Parr JF, Sullivan LA, Quirk R (2009) Sugarcane phytoliths: encapsulation and sequestration of a long-lived carbon fraction. Sugar Tech 11:17–21
- Parr JF, Sullivan LA (2011) Phytolith occluded carbon and silica variability in wheat cultivars. Plant Soil 342:165–171
- Zuo XX, Lü HY (2011) Carbon sequestration within millet phytoliths from dry-farming of crops in China. Chin Sci Bull 56:3451–3456
- Dixon RK, Brown S, Houghton RA et al (1994) Carbon pools and flux of global forest ecosystems. Science 263:185–189
- Malhi Y, Baldocchi DD, Jarvis PG (1999) The carbon balance of tropical, temperate and boreal forests. Plant Cell Environ 22:715–740
- 20. Cao ZH, Zhou GM, Wong MH (2011) Special issue on bamboo and climate change in China. Bot Rev 77:188–189
- 21. Pan Y, Birdsey RA, Fang J et al (2011) A large and persistent carbon sink in the world's forests. Science 333:988–993
- Jiang PK, Meng CF, Zhou GM et al (2011) Comparative study of carbon storage in different forest stands in subtropical China. Bot Rev 77:242–251
- Xu Y, Wong MH, Yang JL et al (2011) Dynamics of carbon accumulation during the fast growth period of bamboo plant. Bot Rev 77:287–295
- 24. Wu ZY, Raven PH (2006) Flora of China: Poaceae. Science Press, Beijing
- 25. Guo QY, Yang GY, Du TZ et al (2005) Carbon character of Chinese bamboo forest. World Bamboo Rattan 3:25–28
- Ceotto E (2008) Grasslands for bioenergy production. Agron Sustain Dev 28:47–55

- Benbi DK, Brar JS (2009) A 25-year record of carbon sequestration and soil properties in intensive agriculture. Agron Sustain Dev 29:257–265
- 28. Li R, Zhang J, Zhang ZE (2003) Values of bamboo biodiversity and its protection in China. J Bamboo Res 22:7–17 (in Chinese)
- Chen LZ, Huang JH, Yan CR (1997) Nutrient cycles in forest ecosystems of China. China Meteorological Press, Beijing (in Chinese)
- Norris AR, Hackney CT (1999) Silica content of a mesohaline tidal marsh in North Carolina. Estuar Coast Shelf Sci 49:597–605
- Motomura H, Mita N, Suzuki M (2002) Silica accumulation in long-lived leaves of *Sasa veitchii* (Carrière) Rehder (Poaceae– Bambusoideae). Ann Bot-London 90:149–152
- 32. Lu RK (2000) Soil agricultural chemical analysis method. Chinese Agricultural Scientific Press, Beijing (in Chinese)
- Parr JF, Dolic V, Lancaster G et al (2011) A microwave digestion method for the extraction of phytoliths from herbarium specimens. Rev Palaeobot Palyno 116:203–212
- 34. Walkley A, Black IA (1934) An examination of the Degtjareff method for determining soil organic matter, and a proposed modification of the chromic acid titration method. Soil Sci 37:29–38
- 35. Murphy DB (2012) Fundamentals of light microscopy and electronic imaging. Wiley, New York
- Kröger N, Lorenz S, Brunner E et al (2002) Self-assembly of highly phosphorylated silaffins and their function in biosilica morphogenesis. Science 298:584–586
- Ding TP, Zhou JX, Wan DF et al (2008) Silicon isotope fractionation in bamboo and its significance to the biogeochemical cycle of silicon. Geochim Cosmochim Acta 72:1381–1395
- Matichenkov V, Calvert D, Snyder G (1999) Silicon fertilizers for citrus in Florida. Proc Fla State Hortic Soc 112:5–8
- Alvarez J, Datnoff LE (2001) The economic potential of silicon for integrated management and sustainable rice production. Crop Prot 20:43–48
- 40. Ma JF, Takahashi E (2002) Soil, fertilizer, and plant silicon research in Japan. Elsevier, Amsterdam
- Mecfel J, Hinke S, Goedel WA et al (2007) Effect of silicon fertilizers on silicon accumulation in wheat. J Plant Nutr Soil Sci 170:769–772
- 42. Zhang YL, Yu L, Liu MD et al (2008) Silicon liberation characteristics of soil and its effect factors after applying slag mucks I relationships between calcium, magnesium, iron and aluminium and silicon liberation. Chin J Soil Sci 39:722–725 (in Chinese)
- 43. Zhao SL, Song ZL, Jiang PK et al (2012) Fractions of silicon in soils of intensively managed *Phyllostachys Pracecox* stands and their plant-availability. Acta Pedol Sin 49:331–338 (in Chinese)
- 44. Qiu EF, Chen ZM, Zheng YS et al (2005) Dynamics of litterfall and its decomposition and nutrient return of shoot-used *Dendrocalamus latiflorus* in Mountainous areas of Fujian Province. Chin J Appl Ecol 16:811–814 (in Chinese)
- 45. Rong JD, Wang XT, Guo XJ et al (2007) Study on the litter and nutrient dynamics of *Dendrocalamus minor* plantation in the costala sandy site. J Fujian Forest Sci Technol 12:59–62 (in Chinese)
- 46. Yu Y, Fei SM, He YP et al (2005) The structure and aboveground biomass of *Pleioblastus amarus* population in Changning. J Sichuan Forest Sci Technol 8:90–93 (in Chinese)
- Zhou GM, Wu JS, Jiang PK (2006) The impacts of different management modes on the carbon storage within moso bamboo. J Beijing For Univ 28:51–55 (in Chinese)
- Hong CT, Fang J, Jin AW et al (2011) Comparative growth, biomass production and fuel properties among different perennial plants, bamboo and miscanthus. Bot Rev 77:197–207
- 49. Ma NX (2004) Resources of sympodial bamboos in China and their utilization. J Bamboo Res 23:1–5 (in Chinese)
- 50. Dong WY (2000) Studies on the high-effective raising seeding technique of scattered bamboo. Tech Rep (in Chinese)