

Variations in nitrogen, zinc, and sugar concentrations in Chinese fir seedlings grown on shrubland and plowed soils in response to arbuscular mycorrhizae-mediated process

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Received: 15 January 2010 / Revised: 30 December 2010 / Accepted: 3 January 2011 / Published online: 13 January 2011
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Abstract One-year-old seedlings of Chinese fir (*Cunninghamia lanceolata* (Lamb.) Hook) are not colonized with ectomy-corrhizal (EM) fungi, but often colonized with arbuscular mycorrhizal (AM) fungi. The AM fungi could be important for nutrient acquisition in Chinese fir seedlings. Previous studies show that feedbacks between above-ground and belowground plant tissues play a fundamental role in controlling the interaction between plants and AM fungi. Our results indicate significant feedback in seedlings grown on shrubland soils, but not on plowed soils. The amounts of sugar in fir leaves in the shrubland soils were significantly lower than those in plowed soils. Leaf zinc (Zn) and nitrogen (N) concentrations were significantly higher in seedlings in shrubland soils than in plowed soils. In mycorrhizal seedlings growing in shrubland soils, leaf N:P ratios were significantly higher than those in plowed soils, likely because of enhanced N absorption through AM-mediated process. Leaf N:P ratios in seedlings grown on plowed soils were below the threshold levels, because of low metabolic activity of feedback in AM-mediated process. The results suggested that the presence of feedback between Chinese fir seedlings and AM fungi should be benefit in transplanting Chinese fir seedlings.

Keywords Arbuscular mycorrhizal fungi · Sugar · Zinc · Nitrogen · Feedback · ^{13}C natural abundance · Chinese fir seedlings

Introduction

Chinese fir (*Cunninghamia lanceolata* (Lamb.) Hook) is usually established for timber production, with a rotation time of about 20–25 years (Zhang et al. 2004). Chinese fir has expanded rapidly since 1980s, accounting for 24% of total planted forest areas in China (Guo et al. 2009). Natural forests have been cleared and replaced by monoculture plantations of Chinese fir in southwest China. Traditional reforestation of Chinese fir involves planting of 1- to 2-year-old seedlings produced in nurseries on clear-cut and slash-and-burn sites that previously harbored native evergreen broad-leaved forests in southwest China. At present, most Chinese fir nurseries and plantations are in the second or third rotation on the same sites. Yield decline and site degradation (Zhang et al. 2004) have been ascribed primarily to a decline in soil C and nutrients largely because of slash burning during site preparation (Zhang et al. 2004). On plowed soils, the seedlings sometimes harbor diseases, and symptoms of N deficiency, such as the yellow discoloration of leaves.

The AM symbiosis is important for N and P nutrition of host (Smith and Read 1997), and depends on the host for sugars for the formation, maintenance, and function of the fungal structures (Zhu and Miller 2003; Ferrol and Pérez-Tienda 2009). Since sucrose is the major form of photo-assimilates transported in higher plants, this sugar should play an important role in C delivery to the symbiosis (Ferrol and Pérez-Tienda 2009). The amount of C allocated to AM is estimated to range from 4% to 20% of a plant's total C budget (Smith and Read 1997). Arbuscular mycorrhiza may improve plant performance when plant is attacked by pathogens or by insect herbivores (Gange 2007). Arbuscular mycorrhiza colonization has been shown

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to enhance plant growth and survival at low soil pH (Clark 1997), and its colonization plays an important role in plant NO_3^- -N uptake (Azcón et al. 2008). However, agricultural practices and clear-felling reduce the abundance and diversity of mycorrhizal fungi (Hedlund and Gormsen 2002; Brundrett 2009), and the destruction of mycorrhizae can lead to serious reforestation problems (Marshall 2000), especially for Chinese Fir. The role of mycorrhizal fungi associated with Chinese fir has not been recognized by farmers and managers. However, an attempt of scaling up and bridging microbial processes to the landscape and regional level can involve the selection of the right microbial indicators (Theuerl and Buscot 2010).

Arbuscular mycorrhiza can also take up Zn, and transfer it to the host plant (Cavagnaro et al. 2006), thereby enhancing plant Zn nutrition, particularly when it is present at low concentrations in soil (Davis et al. 2007). However, under high soil Zn concentrations, the formation of AM can also “protect” against the accumulation of Zn in plant tissues (Cavagnaro 2008). Arbuscular mycorrhiza may also influence nutrient availability via their effects on soil physiochemical and biological properties, such as pH (Li and Christie 2001), and nutrient cycling (Jackson et al. 2008). Plant N and P status can also have a significant effect on plant Zn nutrition (Marschner 1995), and the effects of N and P on AM may have consequences for plant Zn nutrition (Cavagnaro 2008). Despite the fact that numerous scientists have indicated that AM fungi promote N or immobile nutrient Zn of host plants independently, the interaction between N and Zn has rarely been investigated in Chinese fir mycorrhizal system.

In order to investigate the huge differences in Chinese fir seedling growth observed between shrub/grasslands and plowed soils, we divided nurseries into two groups: the first included land supporting shrub/grass after slash burning (4 sites) and sites with one rotation of plantations established after felling native forests (9 sites); the second was >2 rotation plantation sites (7 sites) and plowed sites, mainly paddy soil (4 sites). We measured colonization of mycorrhizal fungi and C isotope composition in seedlings grown on shrub/grass and deciduous forest after slash burning, which were optimum lands for growing seedlings, and grown on the plowed soils and >2 rotation plantation site. As a result of greater discrimination, the photosynthetic C sugars transferred from leaves to roots are more depleted in ^{13}C than nonphotosynthetic C derived from oxalacetate synthesized from phosphoenolpyruvate (PEP) reacting with HCO_3^- in roots, indicating indirectly the effects of AM fungi on plant C isotope composition since they mediate sugar transfer from leaf to root tissues. The aim was to determine whether Chinese fir seedlings grown on different land use histories responded differentially as determined by sugar and Zn concentrations, $\delta^{13}\text{C}$ values, and N:P, and to

determine if N cycling in those plant–soil systems was tightly regulated by AM-mediated process. We hypothesized that despite the degree of colonization level by AM fungi, occurrence of feedbacks between plant tissues should enhance the ability of nutrient acquisition, such as Zn and N for Chinese fir seedlings.

Materials and methods

Research site

In this study, 24 nursery fields on slopes at different elevations above sea level were chosen as the sampling sites. The sites were in mountainous terrain, a transitional zone from low land to hills in the Guizhou province of southwest China. Sampling site elevation ranged from 280 to 1,510 m. Mean annual air temperatures decreased with altitude, and ranged from 18°C to 14.3°C. Most parts of the Guizhou Province have total annual precipitation >1,100 mm. Sampling sites were located in sandstone areas except for one site. The distribution of soil types varies with elevations; yellowish-red soils (Ultisols) were developed at altitudes below 600 m above sea level, and yellow soils (Ultisols) were developed at the altitudes of 600–1,500 m (Piao et al. 2001). The corresponding soil samples were taken at 0–25 cm depth with more than ten replicated cores for each soil sample to provide an indication of nutrient availability. The seedlings of Chinese fir were collected from nurseries in early January 2008 and 2009. The total biomass of each seedling was measured, and it was then separated into leaves, stems, and roots. Site selection aimed to capture a significant range of the variability in soil with usage histories. Fertilizers (15% N, 8% K_2O , 15% P_2O_5) were added in soil surface at all 24 sites before sowing seeds. Plowing was carried out in early spring, at a tillage depth of about 25 cm. Generally, the farmers sow seeds in February, and remove weeds at regular intervals. The nurseries on the shrub/grass and some of the one-rotation sites did not receive fertilizer additions of urea, but those from the plowed soils and >2-year rotation sites had urea added one or two times during one-grown period.

Laboratory analysis

Plant samples for laboratory analysis were dried for 48 h at 60°C and ground with a mortar and a pestle. Total C and N were determined with a CHNS autoanalyzer (PE 2400-II). Metal elements and P were digested using nitric–perchloric acid digestion and analyzed by the atomic absorption spectrometry (AAS) and vanadomolybdate colorimetric methods, respectively. For soluble sugar determination, 0.25 g of air-dried plant material was collected in ten plots

and extracted four times with distilled water at 75°C, modified from the method of Chinnasamy and Bal (2003) because water was used instead of 80% of ethanol and the temperature of boiling water bath was 75°C. After each extraction, samples were filtered (Whatman No 42 filter paper), and the filtrates were used to determine soluble sugar colorimetrically through anthrone reaction (Piao et al. 2000). The concentration of available Zn and Fe of plant material was determined by ASS after diethylenetriamine-pentaacetic acid (DTPA) extraction (Lindsay and Norvell 1978). Standard soil analysis methods were used to measure soil-extractable Ca and Mg with 1 M NH₄OAC (Thomas 1982). The $\delta^{13}\text{C}$ values of plants were measured by combustion of 2-mg homogenized samples with CuO (1:50) at 850°C in a vacuum-combustion system. Carbon dioxide generated in the combustion tubes was separated by cryogenic distillation, collected in breakseals and analyzed on a mass spectrometer (MAT 252). The samples of Chinese fir seedlings were sent to the laboratory of China Agricultural University and examined for mycorrhizal colonization following the method of Gai et al. (2006). Briefly, the cleared roots with 10% KOH were acidified in lactic acid, and stained with Trypan blue, and examined for colonization level under a compound microscope. The data for $\delta^{13}\text{C}$ are expressed relative to the international standard PDB (as $\delta^{13}\text{C}$ ‰ (parts per thousand)). Each sample was analyzed in duplicate. The standard errors of C and N concentrations were always less than 0.01 g C kg⁻¹, and the standard deviations of the C isotope composition did not exceed 0.1‰. Differences between mean values of soil and plant nutrient concentrations were tested for their significance by using linear regression and variance analysis at 5% of probability.

Results

All of Chinese fir seedlings collected here were colonized by AM fungi, but not by EM fungi. Colonization levels with AM fungi varied from <10% to 74% with no differences between shrubland and plowed soils. According to field investigations, all of Chinese fir seedlings grown on shrubland soils did not harbor root rot and aboveground disease, but some of seedlings grown on plowed soils did, however, the cause the disease and N deficiency symptoms.

There were no significant differences in soil properties between shrubland and plowed soils. The average soil pH was 4.40±0.28, ranging from 4.08 to 4.96 in shrub soils, and 4.80±0.72, ranging from 4.18 to 5.14, with exception for one of 6.89, in plowed soil. The exchangeable Ca and Mg concentrations were 0.51±0.34 mg g⁻¹ and 0.049±0.022 mg g⁻¹ in shrub soils, and were 0.75±0.76 mg g⁻¹

and 0.10±0.19 mg g⁻¹ in plowed soils, respectively. The average ratio of soil C:N (13.0±2.3) in shrubland soils was similar to that in plowed soils (12.9±2.6). Available Zn extracted with DTPA solution (1.9±0.9 μg g⁻¹) in shrub soils was similar to that (1.7±0.9 μg g⁻¹) in plowed soils, and iron (Fe) concentration (0.080±0.029 mg g⁻¹) in shrub soils was also similar to that (0.073±0.042 mg g⁻¹) in plowed soils.

Most parameters of leaf tissues did not statistically differ between shrub and plowed soils (Table 1); however, the average concentrations of leaf sugar ranged from 18 to 166 mg g⁻¹ in shrub soils and were significantly lower ranging from 56 to 164 mg g⁻¹ in plowed soils. Leaf Zn ranging from 3 to 45 μg g⁻¹ in shrub soils was significantly higher than values ranging from 3 to 26 μg g⁻¹ in plowed soils. The leaf N:P ranging from 11.6 to 17.1 (mass) in shrubland soils was significantly higher than in plowed soils, where N:P ranged from 6.6 to 13.2 (mass; Table 1). Leaf C (%) ranging from 47.2% to 51.0% in shrub was higher than that in plowed soils, where leaf C ranged from 46.7% to 49.7%. Among root tissues, only Zn significantly differed between shrub and plow soils (Table 1).

Although there was no significant correlation between leaf sugar and biomass (height; $r^2=0.024$, $P>0.05$), biomass significantly decreased by decreasing root sugar ($r^2=0.385$, $P<0.01$; Fig. 1a). Leaf N was correlated positively with leaf P ($r^2=0.311$, $P<0.01$), and significantly correlated with leaf Zn ($r^2=0.413$, $P<0.001$; Fig. 1b). Leaf Zn significantly decreased with increasing leaf sugar ($r^2=0.175$, $P<0.05$), and leaf N also decreased with increasing leaf sugar ($r^2=0.212$, $P<0.05$; Fig. 1c). It seems that there was metabolic feedback between aboveground–belowground tissues in seedlings grown on shrubland soils. Leaf Zn was not significantly correlated with leaf P ($r^2=0.003$). Therefore, the significant correlation between leaf Zn and leaf N:P ($r^2=0.198$, $P<0.05$) was mainly determined by the variation of leaf N. Root N was not significantly correlated with biomass (height; $r^2=0.009$) and root sugar ($r^2=0.028$), respectively, whereas the root Zn was negatively and significantly correlated with biomass ($r^2=0.293$, $P<0.01$; Fig. 1d) and root sugar ($r^2=0.403$, $P<0.001$). This relationship was similar in fir seedlings on shrubland and plowed soils. This may suggest that the concentrations of sugar in the root did not necessarily reflect the degree to which sugar was available to the fungus.

The leaf biomass was always ¹³C-depleted (−28.5±0.5‰) relative to the root tissues (−26.9±0.6‰). There was no difference of leaf $\delta^{13}\text{C}$ between seedlings from shrubland (−28.4±0.4‰) and plowed sites (−28.7±0.5‰), whereas root $\delta^{13}\text{C}$ (−26.7±0.5‰) in shrub was higher than that (−27.1±0.6‰) in plow.

Table 1 Some chemical properties of Chinese fir seedlings

Tissue		C (%)	$\delta^{13}\text{C}$ (‰)	Zn ($\mu\text{g g}^{-1}$)	Sugar (mg g^{-1})	Ca (mg g^{-1})	Mg (mg g^{-1})	Fe (mg g^{-1})	N (mg g^{-1})	P (mg g^{-1})	N:P ratio (mass)
Leaf	Shrub	48.7 (0.9)	-28.4 (0.4)	25.1 (10.9)	85 (36)	7.5 (1.4)	1.13 (0.17)	0.11 (0.04)	20.8 (4.1)	1.5 (0.3)	14.2 (1.6)
	Plow	47.9 (0.7)	-28.7 (0.3)	15.7 (6.7)	124 (30)	7.5 (1.4)	1.20 (0.36)	0.11 (0.01)	17.4 (3.7)	1.7 (0.5)	10.8 (1.9)
	<i>t</i> test	$P < 0.05$	$P > 0.05$	$P < 0.01$	$P < 0.01$	$P > 0.05$	$P > 0.05$	$P > 0.05$	$P > 0.05$	$P > 0.05$	$P > 0.05$
Root	Shrub	45.8 (1.6)	-26.7 (0.5)	21.3 (11.3)	53.2 (22.1)	2.5 (0.4)	1.2 (0.4)	0.37 (0.14)	8.4 (2.1)	0.9 (0.2)	10.1 (2.5)
	Plow	46.4 (2.5)	-27.1 (0.6)	12.6 (9)	72.1 (30.7)	3.0 (0.5)	1.3 (0.5)	0.41 (0.23)	8.8 (4.0)	1.2 (0.5)	8.3 (5.8)
	<i>t</i> test	$P > 0.05$	$P > 0.05$	$P < 0.05$	$P > 0.05$	$P > 0.05$	$P > 0.05$	$P > 0.05$	$P > 0.05$	$P > 0.05$	$P > 0.05$

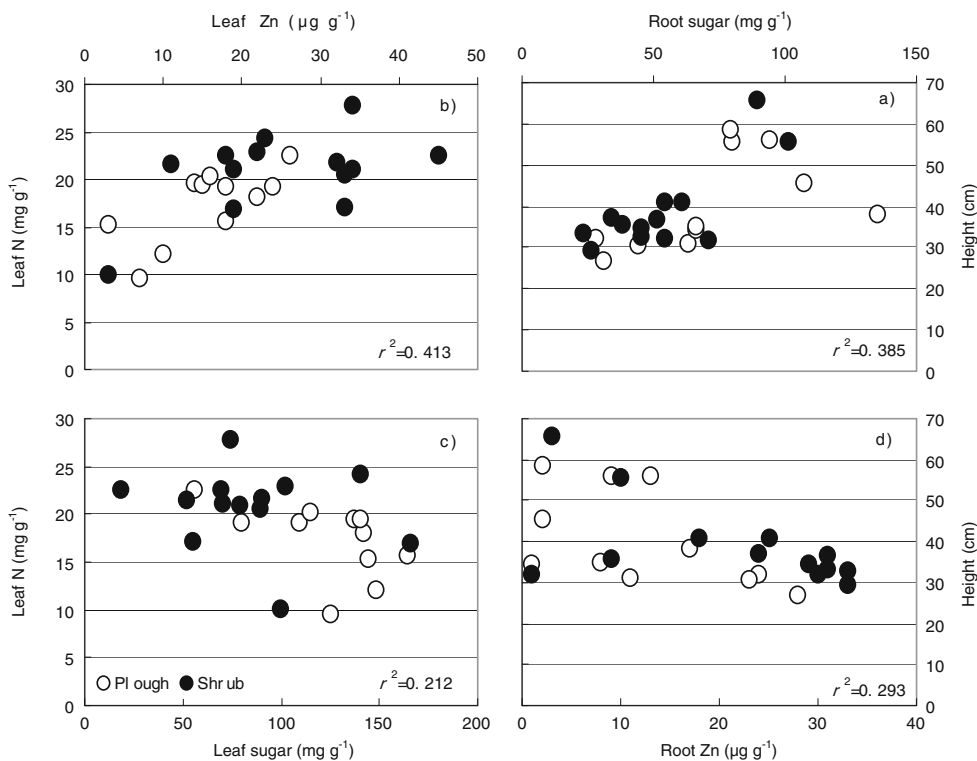
Discussion

Our results showed that there was no significant relationship between AM colonization levels and nutrient acquisitions in seedlings grown on both shrubland and plowed soils. But there were significant differences in the internal feedbacks in fir seedlings on shrubland and plowed soils. A key AM-mediated process is the transfer of photosynthate from host plant to AM fungi hyphae, and its residence time is normally days (Zhu and Miller 2003; Richardson et al. 2009). The fungi are obligate symbionts and cannot survive without photosynthate supply from plants (Smith et al. 2010). When less C is transferred from plants to fungus, this may be responsible for low concentrations of nutrients in seedling tissues grown on plowed soils. The extraradical hyphae of the AM fungi are a key component of AM symbiosis linking colonized roots with the soil matrix (Malcová et al. 2001),

proliferating in the surrounding soil from which they absorb nutrients (Hawkins and George 2001). Our results suggested that colonized roots may play a more important role in exploiting N in plowed soils than in shrub/grassland soils. Soil disturbance and high soil compaction can destroy hyphal networks and reduce AM fungal populations (Drew et al. 2006), and mineral fertilization can reduce N uptake by AM fungi hyphae and mycorrhizal roots (Cheng et al. 2008). However, inorganic fertilizer was added in all of 24 sites onetime. It has been shown that urea does not affect on soil microbiological properties (Lupwayi et al. 2010).

At the sampling time, we observed new root emergence in shrubland sites, but rarely in plowed sites. The intraradical colonization by mycorrhizal fungi is generally faster (peaking at 7 to 15 days) than that by nonmycorrhizal fungi and often more frequently in younger roots (Resendes et al. 2008). We found that even the seedlings showing N-

Fig. 1 Aboveground height as affected by root sugar (a), N content of leaf as affected by Zn (b) or sugar content of leaf (c), and aboveground height as affected by root Zn (d)



deficient symptoms (collected at January 2010) were also colonized by AM fungi (up to 36%) and developed root hair in sites after several rotations. Mycorrhizae are particularly important for nutrient uptake in plant species that do not develop dense root systems or do not develop root hairs (Jakobsen et al. 2005). It is apparent that roots grown on shrubland soils had higher metabolic activity than those grown on plowed soils, which should be responded to different soil conditions.

Aboveground–belowground feedbacks play a fundamental role in controlling the interaction between plants and soil organisms including AM fungi (Wardle et al. 2004). As already mentioned, the fungi efficiently take up inorganic ions from the soil and transfer them to the plant (Nygren et al. 2007), while the plant supplies the mycorrhizal fungi with products of photosynthesis (Baum et al. 2009), and the C transport is improved by root colonization by AM fungi. The low sugar in leaf tissues was correlated with higher leaf N and Zn in seedlings grown on shrubland soils in comparison with plowed soils, indicating that there was apparent aboveground–belowground feedback regulation. The lower leaf sugar in shrubland than in plowed soils should imply that more sugar was transferred to AM fungi-mediated processes in the former than in the latter soil, and extraradical hyphae of AM more efficiently absorb N and Zn in the former soils, and these nutrients were transferred from root to leaf tissues, resulting in increasing contents of leaf N and Zn in plants grown on shrubland soils. Previous study also shows that there is significantly less sucrose in both shoot and root in summer wheat with AM fungus compared to those without AM fungus (Hawkins and George 2001).

Threshold levels for nutrients in plant tissues have been suggested by Koerselman and Meuleman (1996), who indicate that $N:P > 16$ indicates P limitation and $N:P < 13$ shows N limitation. Characteristic ratios between N and P determine the basic stoichiometry for the plants, allowing us to predict their response to alterations of N and P availability (Schlesinger 2004). The seedlings grown on shrubland soils had higher N:P ratio values than threshold levels, except for 2 out of 13 sites, and seedlings on plowed soils had the N:P ratio values lower than threshold levels, except for 2 out of 11 sites. Therefore, most sites of plowed soils but not those of shrubland soils were under N-limited conditions. In the N-limited soils, like our nursery sites, the N delivery to the host plant from the mycorrhizal fungi depends on the C flow from the plant (Egerton-Warburton et al. 2007; Baum et al. 2009), and the transfer of plant C to mycorrhizal fungi is higher under conditions of low nutrient supply (Huygens et al. 2008).

It has long been thought that AM fungi do not contribute significantly to nitrate uptake by plants, but Azcón et al. (2008) demonstrated that NO_3^- appeared to be the best N

source for AM plants in a neutral-alkaline soil and particularly under drought stress conditions. Chinese fir normally grows on low pH, moist, and fertile soils. Soil pH is known to have large effects on N cycling (Ste-Marie and Paré 1999). Despite the decrease in biological activities, net mineralization of N and P increases by decreasing pH (Kooijman and Hedenäs 2009). There are constraints on nitrification other than substrate supply in low soil pH, with N-limited conditions (Falkengren-Grerup et al. 2004). Net nitrate production in acidic forest floor is often very low or clearly absent, with prevalence of NH_4^+ (Ste-Marie and Paré 1999); even so, nitrate may still be a significant proportion of the N taken up by trees (George et al. 1999). By determining ^{15}N natural abundance, we have found that the increases of leaf N in seedlings grown on shrubland soils were associated with absorption of nitrate by Chinese fir (nonpublished data).

As already mentioned, AM can also increase uptake of Zn (Thompson 1990; Purakayastha and Chhonkar 2001; Cavagnaro et al. 2006). The contribution of mycorrhiza to total Zn uptake is estimated at approximately 50% in clover (Marschner 1995). Arbuscular mycorrhizal fungi may also influence nutrient availability via their effects on soil physiochemical properties, like soil pH (Li and Christie 2001), and nutrient cycling (Jackson et al. 2008). In addition, acidification of rhizosphere soil can improve Zn solubility and thus Zn availability (Cavagnaro 2008). Preferential solubilization of highly insoluble fractions, such as crystalline-bound and residual fractions of Zn, by AM fungi can occur in soil (Subramanian et al. 2009). Probably, the observed increase in Zn uptake by seedlings grown on shrubland soils was associated with higher Zn solubilization than in plowed soils.

The leaf biomass was always ^{13}C -depleted relative to the root, due to either fractionation processes or an increased proportion of PEPc fixed C in root tissues (Brandes et al. 2006). In our case, no differences of leaf $\delta^{13}C$ were found between shrubland and plowed sites. The root $\delta^{13}C$ in fir seedlings grown on shrubland soils was higher relative to that in plowed soils. Roots can take up HCO_3^- and consume it through the root PEPc. The selective incorporation is higher in roots than in leaves (Ford et al. 2007), and approximately 1% of total plant C can originate from root uptake of inorganic C (Ford et al. 2007; Rasmussen 2009). The fact that PEPc enzymatic activity is higher in NO_3^- -fed than NH_4^+ -fed plants could be explained by alkalinization of cellular sap caused by the reduction of the taken up nitrate in root cells (Pasqualini et al. 2001). However, the NH_4^+ nutrition can enhance the activity of PEPc in numerous plant species, and thus the bicarbonate assimilation in root cells (Pasqualini et al. 2001). In our case, the assimilations of HCO_3^- in the roots grown on shrubland soils may be higher than in plowed soils. In

addition, the difference of root $\delta^{13}\text{C}$ between shrubland and plowed soils could also be ascribed to difference in the amount of photosynthates delivered to the AM-infected roots.

Conclusion

To avoid parasitism, plants appear to develop feedback mechanisms to regulate the C drain to the fungal symbiont in relation to the nutrients given by fungus. Chinese fir seedlings are generally transplanted at early January every year. Because of key ecological functions performed by AM associations, low metabolic activity of feedback in AM-mediated process, and loss of mycorrhizal activity in degraded areas may limit the successful re-establishment of Chinese fir seedlings. At present, the information on mycorrhizal colonization in Chinese fir nursery and plantations is scarce, and an intensive sampling is required to resolve the mycorrhizal status of Chinese fir plant.

Acknowledgments We thank Prof. William H. Schlesinger, the Cary Institute of Ecosystem Studies, for assistance in revising the manuscript, and Dr. J. P. Gai, Department of Plant Nutrition, China Agricultural University, for determining the colonization with mycorrhizal fungi. This study was financially supported by the National Natural Science Foundation of China (grant no. 40772207), and by the Ministry of Science and Technology of China (grant no. 2006CB403200).

References

- Azcón R, Rodríguez R, Amora-Lazcano E, Ambrosano E (2008) Uptake and metabolism of nitrate in mycorrhizal plants as affected by water availability and N concentration in soil. *Eur J Soil Sci* 59:131–138. doi:10.1111/j.1365-2389.2007.00962.x
- Baum C, Toljander YK, Eckhardt K-U, Weih M (2009) The significance of host-fungus combinations in ectomycorrhizal symbioses for the chemical quality of willow foliage. *Plant Soil* 323:213–224. doi:10.1007/s11104-009-9928-x
- Brandes E, Kodama N, Whittaker K, Weston C, Rennenberg H, Keitel C, Adams M, Gessler A (2006) Short-term variation in the isotopic composition of organic matter allocated from the leaves to the stem of *Pinus sylvestris*: effects of photosynthetic and postphotosynthetic carbon isotope fractionation. *Global Change Biol* 12:1922–1939. doi:10.1111/j.1365-2486.2006.01205.x
- Brundrett MC (2009) Mycorrhizal associations and other means of nutrition of vascular plants: understanding the global diversity of host plants by resolving conflicting information and developing reliable means of diagnosis. *Plant Soil* 320:37–77. doi:10.1007/s11104-008-9877-9
- Cavagnaro TR (2008) The role of arbuscular mycorrhizas in improving plant zinc nutrition under low soil zinc concentrations: a review. *Plant Soil* 304:315–325. doi:10.1007/s11104-008-9559-7
- Cavagnaro TR, Jackson LE, Six J, Ferris H, Goyal S, Asami D, Scow KM (2006) Arbuscular mycorrhizas, microbial communities, nutrient availability, and soil aggregates in organic tomato production. *Plant Soil* 282:209–225. doi:10.1007/s11104-005-5847-7
- Cheng XM, Euliss A, Baumgartner K (2008) Nitrogen capture by grapevine roots and arbuscular mycorrhizal fungi from legume cover-crop residues under low rates of mineral fertilization. *Biol Fertil Soils* 44:965–973. doi:10.1007/s00374-008-0281-7
- Chinnasamy G, Bal AK (2003) Seasonal changes in carbohydrates of perennial root nodules of beach pea. *J Plant Physiol* 160:1185–1192
- Clark RB (1997) Arbuscular mycorrhizal adaptation, spore germination, root colonization, and host plant growth and mineral acquisition at low pH. *Plant Soil* 192:15–22
- Davis MR, Coker G, Parfitt RL, Simcock R, Clinton PW, Garrett LG, Watt MS (2007) Relationships between soil and foliar nutrients in young densely planted mini-plots of *Pinus radiata* and *Cupressus lusitanica*. *Forest Ecol Manage* 240:122–130. doi:10.1016/j.foreco.2006.12.023
- Drew EA, Murray RS, Smith SE (2006) Functional diversity of external hyphae of AM fungi: ability to colonise new hosts is influenced by fungal species, distance and soil conditions. *Appl Soil Ecol* 32:350–365. doi:10.1016/j.apsoil.2005.07.005
- Egerton-Warburton LM, Querejeta JI, Allen MF (2007) Common mycorrhizal networks provide a potential pathway for transfer of hydraulically lifted water between plants. *J Exp Bot* 58:1473–1483. doi:10.1093/jxb/erm009
- Falkengren-Grerup U, Michelsen A, Olsson MO, Quarmby C, Sleep D (2004) Plant nitrate use in deciduous woodland: the relationship between leaf N, ^{15}N natural abundance of forbs and soil N mineralisation. *Soil Biol Biochem* 36:1885–1891. doi:10.1016/j.soilbio.2004.05.009
- Ferrol N, Pérez-Tienda J (2009) Coordinated nutrient exchange in arbuscular mycorrhiza. In: Azcón-Aguilar C et al. (eds) *Mycorrhizas-functional processes and ecological impact*. doi:10/1007-978-3-540-87978-7_6, Springer, Berlin
- Ford CR, Wurzbarger N, Hendrick RL, Teskey RO (2007) Soil DIC uptake and fixation in *Pinus taeda* seedlings and its C contribution to plant tissues and ectomycorrhizal fungi. *Tree Physiol* 27:375–383
- Gai JP, Feng G, Cai XB, Christie P, Li XL (2006) A preliminary survey of the arbuscular mycorrhizal status of grassland plants in southern Tibet. *Mycorrhiza* 16:191–196. doi:10.1007/s00572-005-0032-7
- Gange AC (2007) Insect-mycorrhizal interactions: patterns, processes, and consequences. In: Price PW, Timothy PC (eds) *Ecological communities: plant mediation in indirect interaction webs*. Cambridge University Press, New York
- George E, Stober C, Seith B (1999) The use of different soil nitrogen sources by young Norway spruce plants. *Trees* 13:199–205
- Guo J-F, Yang Y-S, Liu L-Z, Zhao Y-C, Chen Z-W, Mao Y-L (2009) Effect of temperature on soil respiration in a Chinese fir forest. *J Forest Res* 20:49–53. doi:10.1007/s11676-009-0009-z
- Hawkins H-J, George E (2001) Reduced ^{15}N -nitrogen transport through arbuscular mycorrhizal hyphae to *Triticum aestivum* L. supplied with ammonium vs. nitrate nutrition. *Ann Bot* 87:303–311. doi:10.1006/anbo.2000.1305
- Hedlund K, Gormsen D (2002) Mycorrhizal colonization of plants in set-aside agricultural land. *Appl Soil Ecol* 19:71–78
- Huygens D, Deneff K, Vandeweyer R, Godoy R, Van Cleemput O, Boeckx P (2008) Do nitrogen isotope patterns reflect microbial colonization of soil organic matter fractions? *Biol Fertil Soils* 44:955–964. doi:10.1007/s00374-008-0280-8
- Jackson LE, Burger M, Cavagnaro TR (2008) Roots, nitrogen transformations, and ecosystem services. *Annu Rev Plant Biol* 59:341–363. doi:10.1146/annurev.arplant.59.032607.092932
- Jakobsen I, Chen B, Munkvold L, Lundsgaard T, Zhu YG (2005) Contrasting phosphate acquisition of mycorrhizal fungi with that of root hairs using rootless barley mutant. *Plant Cell Environ* 28:928–938

- Koerselman W, Meuleman AFM (1996) The vegetation N:P ratio: a new tool to detect the nature of nutrient limitation. *J Appl Ecol* 33:1441–1450
- Kooijman A, Hedenäs L (2009) Changes in nutrient availability from calcareous to acid wetland habitats with closely related brown moss species: increase instead of decrease in N and P. *Plant Soil* 324:267–278. doi:10.1007/s11104-009-9954-8
- Li XL, Christie P (2001) Changes in soil solution Zn and pH and uptake of Zn by arbuscular mycorrhizal red clover in Zn-contaminated soil. *Chemosphere* 42:201–207
- Lindsay WL, Norvell WA (1978) Development of a DTPA test for zinc, iron, manganese, and copper. *Soil Sci Soc Am J* 42:421–428. doi:10.2136/sssaj1978.03615995004200030009x
- Lupwayi NZ, Grant CA, Soon YK, Clayton GW, Bittman S, Malhi SS, Zebbarth BJ (2010) Soil microbial community response to controlled-release urea fertilizer under zero tillage and conventional tillage. *Appl Soil Ecol* 45:254–261. doi:10.1016/j.apsoil.2010.04.013
- Malcová R, Albrechtová J, Vosátka M (2001) The role of the extraradical mycelium network of arbuscular mycorrhizal fungi on the establishment and growth of *Calamagrostis epigejos* in industrial waste substrates. *Appl Soil Ecol* 18:129–142
- Marschner H (1995) Mineral nutrition of higher plants, 2nd edn. Academic, London
- Marshall VG (2000) Impacts of forest harvesting on biological processes in northern forest soils. *Forest Ecol Manage* 133:43–60
- Nygren CMR, Edqvist J, Elfstrand M, Heller G, Taylor FS (2007) Detection of extracellular protease activity in different species and genera of ectomycorrhizal fungi. *Mycorrhiza* 17:241–248. doi:10.1007/s00572-006-0100-7
- Pasqualini S, Ederli L, Piccioni C, Batini P, Bellucci M, Arcioni S, Antonielli M (2001) Metabolic regulation and gene expression of root phosphoenolpyruvate carboxylase by different nitrogen sources. *Plant Cell Environ* 24:439–447
- Piao HC, Hong YT, Yuan ZY (2000) Seasonal changes of microbial biomass carbon related to climatic factors in soils from kast areas southwest China. *Biol Fertil Soils* 30:294–297
- Piao HC, Liu GS, Wu YY, Xu WB (2001) Relationships of soil microbial biomass carbon and organic carbon with environmental parameters in mountainous soils of southwest China. *Biol Fertil Soils* 33:347–350. doi:10.1007/s003740000328
- Purakayastha TJ, Chhonkar PK (2001) Influence of vesicular-arbuscular mycorrhizal fungi (*Glomus etunicatum* L.) on mobilization of zinc in wetland rice (*Oryza sativa* L.). *Biol Fertil Soils* 33:323–327. doi:10.1007/s003740000330
- Rasmussen J (2009) Carbon isotopes as proof for plant uptake of organic nitrogen: relevance of inorganic carbon uptake. *Soil Biol Biochem* 41:1586–1587. doi:10.1016/j.solilbio.2009.03.006
- Resendes ML, Bryla DR, Eissenstat DM (2008) Early events in the life of apple roots: variation in root growth rate is linked to mycorrhizal and nonmycorrhizal fungal colonization. *Plant Soil* 313:175–186. doi:10.1007/s11104-008-9690-5
- Richardson AE, Barea J-M, McNeill AM, Prigent-Combaret C (2009) Acquisition of phosphorus and nitrogen in the rhizosphere and plant growth promotion by microorganisms. *Plant Soil* 321:305–339. doi:10.1007/s11104-009-9895-2
- Schlesinger WH (2004) Better living through biogeochemistry. *Ecology* 85:2402–2407
- Smith SE, Read DJ (1997) Mycorrhizal symbiosis. 2nd edn. Academic Press, San Diego, CA
- Smith SE, Facelli E, Pope S, Smith FA (2010) Plant performance in stressful environments: interpreting new and established knowledge of the roles of arbuscular mycorrhizas. *Plant Soil* 326:3–20. doi:10.1007/s11104-009-9981-5
- Ste-Marie C, Paré D (1999) Soil, pH and N availability effects on net nitrification in the forest floors of a range of boreal forest stands. *Soil Biol Biochem* 31:1579–1589
- Subramanian KS, Tenshia V, Jayalakshmi K, Ramachandran V (2009) Biochemical changes and zinc fractions in arbuscular mycorrhizal fungus (*Glomus intraradices*) inoculated and uninoculated soils under differential zinc fertilization. *Appl Soil Ecol* 43:32–39. doi:10.1016/j.apsoil.2009.05.009
- Theuerl S, Buscot F (2010) Laccases: toward disentangling their diversity and functions in relation to soil organic matter cycling. *Biol Fertil Soils* 46:215–225. doi:10.1007/s00374-010-0440-5
- Thomas GW (1982) Exchangeable cations. In: Page AL, Miller RH, Keeney (eds) *Methods of soil analysis, Part 2. Chemical and microbiological properties—Agronomy Monograph no. 9* (2nd edn). ASA-SSSA, 677 S, Segoe RD., Madison, WI, pp 159–165
- Thompson JP (1990) Soil sterilization methods to show VA-mycorrhizal aid phosphorus and zinc nutrition of wheat in vertisols. *Soil Biol Biochem* 22:229–240
- Wardle DA, Bardgett RD, Klironomos JN, Setälä H, Van der Putten WH, Wall DH (2004) Ecological linkages between aboveground and belowground biota. *Science* 304:1629–1633
- Zhang X-Q, Kirschbaum MUF, Hou Z-H, Guo Z-H (2004) Carbon stock changes in successive rotations of Chinese fir (*Cunninghamia lanceolata* (Lamb) Hook) plantations. *Forest Ecol Manage* 202:131–147. doi:10.1016/j.foreco.2004.07.032
- Zhu YG, Miller RM (2003) Carbon cycling by arbuscular mycorrhizal fungi in soil-plant systems. *Trends Plant Sci* 8:407–409